

Research



Cite this article: Knickmeyer MD, Mateo JL, Eckert P, Roussa E, Rahhal B, Zuniga A, Krieglstein K, Wittbrodt J, Heermann S. 2018 TGF β -facilitated optic fissure fusion and the role of bone morphogenetic protein antagonism. *Open Biol.* **8**: 170134. <http://dx.doi.org/10.1098/rsob.170134>

Received: 31 May 2017
Accepted: 2 March 2018

Subject Area:
developmental biology

Keywords:
optic fissure fusion, coloboma, TGF β , BMP, ECM

Author for correspondence:
Stephan Heermann
e-mail: stephan.heermann@anat.uni-freiburg.de

[†]Present address: School of Medicine and Health Sciences, An-Najah National University, Nablus, Palestine.

Electronic supplementary material is available online at <https://dx.doi.org/10.6084/m9.figshare.c.4035245>.

TGF β -facilitated optic fissure fusion and the role of bone morphogenetic protein antagonism

Max D. Knickmeyer^{1,2}, Juan L. Mateo³, Priska Eckert^{1,2}, Eleni Roussa¹, Belal Rahhal^{1,†}, Aimee Zuniga⁴, Kerstin Krieglstein¹, Joachim Wittbrodt⁵ and Stephan Heermann¹

¹Department of Molecular Embryology, Institute of Anatomy and Cell Biology, Faculty of Medicine, University of Freiburg, Freiburg D-79104, Germany

²Faculty of Biology, University of Freiburg, Schaezlestrasse 1, Freiburg D-79104, Germany

³Departamento de Informática, Universidad de Oviedo, Jesús Arias de Velasco, Oviedo 33005, Spain

⁴Developmental Genetics, University of Basel Medical School, Basel CH-4058, Switzerland

⁵Centre for Organismal Studies, Heidelberg D-69120, Germany

SH, 0000-0001-7374-8886

The optic fissure is a transient gap in the developing vertebrate eye, which must be closed as development proceeds. A persisting optic fissure, coloboma, is a major cause for blindness in children. Although many genes have been linked to coloboma, the process of optic fissure fusion is still little appreciated, especially on a molecular level. We identified a coloboma in mice with a targeted inactivation of transforming growth factor β 2 (TGF β 2). Notably, here the optic fissure margins must have touched, however failed to fuse. Transcriptomic analyses indicated an effect on remodelling of the extracellular matrix (ECM) as an underlying mechanism. TGF β signalling is well known for its effect on ECM remodelling, but it is at the same time often inhibited by bone morphogenetic protein (BMP) signalling. Notably, we also identified two BMP antagonists among the downregulated genes. For further functional analyses we made use of zebrafish, in which we found TGF β ligands expressed in the developing eye, and the ligand binding receptor in the optic fissure margins where we also found active TGF β signalling and, notably, also gremlin 2b (*grem2b*) and follistatin a (*fsta*), homologues of the regulated BMP antagonists. We hypothesized that TGF β is locally inducing expression of BMP antagonists within the margins to relieve the inhibition from its regulatory capacity regarding ECM remodelling. We tested our hypothesis and found that induced BMP expression is sufficient to inhibit optic fissure fusion, resulting in coloboma. Our findings can likely be applied also to other fusion processes, especially when TGF β signalling or BMP antagonism is involved, as in fusion processes during orofacial development.

1. Introduction

The optic fissure is a transient gap in the developing vertebrate eye. It is an entry route used by cells of the periocular mesenchyme and embryonic vasculature. However, it is necessary that the fissure is closed as development proceeds. A persisting fissure is termed coloboma. A coloboma can affect vision severely and is a frequent cause for blindness in children [1]. Many genes and signalling pathways have been linked to coloboma [2–11], resulting in a growing gene coloboma network [12,13]. Coloboma is frequently part of a multi-organ syndrome, like CHARGE syndrome or renal coloboma syndrome, linked to Chd7 and Pax2, respectively [14–18]. Notably, the morphology of coloboma phenotypes is highly variable. Alterations in some signalling pathways (e.g. Wnt, Hippo) result in vast extended coloboma [4,7], likely originating from early morphogenetic

defects. In this context, we recently linked a precocious arrest of a bilateral neuroretinal flow during optic cup formation to an extended coloboma [11]. There, we found that a locally expressed antagonist to bone morphogenetic proteins (BMPs) was necessary to maintain the tissue flow. Importantly, such massive coloboma phenotypes are morphologically different from coloboma resulting from a hampered fusion process of the optic fissure margins. The pathomechanisms behind such phenotypes, however, are largely elusive, and so is the underlying physiological process. This process is not understood on a structural and especially on a molecular level.

Preceding the fusion, the prospective neuroretina and retinal pigmented epithelium (RPE) share a basement membrane within the optic fissure margin. In order to facilitate the fusion of the margins, the structure of the epithelial margins must be rearranged somehow. This was shown to affect cell–cell connections [19] and likely affects the extracellular matrix (ECM) as well. Concerning cell–cell connections it should be noted that mutants for N-cadherin and α -catenin showed coloboma phenotypes [20,21]. However, it remains unclear at which point during margin disassembly, fusion or consecutive reassembly of the neuroretina and RPE these factors play a role. Although it was shown *per se* that the dissolution of the basement membrane occurs [19] and is a prerequisite for fusion [22], the effector molecules for the structural remodelling and epithelial disassembly, which facilitate the fusion process, are largely elusive.

Transforming growth factor β (TGF β) signalling is well known to induce changes to the ECM and, furthermore, to trigger epithelial to mesenchymal transition in various processes during development and disease [23–28]. Notably, TGF β -regulated changes to the ECM are frequently inhibited by BMP signalling [29–32].

Here we addressed the role of TGF β and BMP in optic fissure fusion, making use of mouse (*Mus musculus*) and zebrafish (*Danio rerio*).

Our findings show that TGF β signalling is acting pro-fusion upon the optic fissure margins. We identified a coloboma phenotype in the TGF β 2 knockout (KO) mouse, with TGF β ligands expressed in the zebrafish eye, and the ligand binding receptor expressed within the fissure where we also found active TGF β signalling. We identified two TGF β -dependent BMP signalling antagonists in mouse, and homologues of these we found expressed within the optic fissure margins in zebrafish. Notably, in zebrafish, both TGF β signalling inhibition and forced BMP expression during fissure fusion are sufficient to prevent optic fissure fusion, resulting in coloboma.

Based on our data we propose that TGF β signalling is locally inducing BMP antagonists to relieve a BMP-induced inhibition on ECM remodelling, eventually allowing TGF β signalling to act pro-fusion.

2. Results

2.1. Loss of TGF β 2 results in coloboma

In the mouse genome, three TGF β isoforms are encoded (TGF β 1, 2 and 3). Targeted inactivation of TGF β 2 results in several phenotypes, also affecting the eye [33], e.g. a remaining primary vitreous, a Peters anomaly like phenotype and an altered neuroretinal layering. In addition to these phenotypes, we identified a persistent optic fissure in TGF β 2 mutant embryos (figure 1b,

electronic supplementary material, figure S1A, B, figure 1a as control). TGF β 2-dependent coloboma was first observed in TGF β 2/GDNF double mutants ([34], Rahhal & Heermann 2009, unpublished observations, electronic supplementary material, figure S1C) and subsequently in TGF β 2 single mutants, derived from the same breeding background (this study, figure 1b, electronic supplementary material figure S1A, B), but not in GDNF single mutants. Furthermore, no phenotype affecting eye development was described in the three distinct GDNF mutant mice [35–37], although GDNF expression was documented in the developing eye [38]. Notably, we found marked, severe coloboma phenotypes in both TGF β 2 KO (figure 1b, electronic supplementary material, figure S1A, B) as well as in TGF β 2/GDNF double KO conditions (electronic supplementary material, figure S1C). Overall, the optic fissure margins in our colobomatous embryos must have been in close proximity to each other, but ultimately failed to fuse and instead grew inwards towards the lens (figure 1b, electronic supplementary material, figure S1A, B, showing different sections of individual eyes). Since we found such coloboma phenotypes in both TGF β 2 and TGF β 2/GDNF mutants, but not in GDNF mutants, we found it likely that the phenotype was resulting from a TGF β 2 loss. A sensitizing role of GDNF in this scenario, however, cannot be ruled out.

So far, we based the analyses on TGF β 2 single mutants from a mixed breeding background [34] (figure 1b, see a as control). We next asked whether this breeding background could have an effect on the analyses. We additionally addressed TGF β 2 mutants derived from a sole background. Notably, while the overall coloboma phenotype was variable in intensity in the TGF β 2 single mutants from a mixed breeding background (e.g. figure 1b, electronic supplementary material, figure S1A, B), we could only detect very subtle forms of coloboma in TGF β 2 single mutants from a sole background (figure 1d, figure 1c as control). This suggests that the breeding background has an effect on the coloboma phenotype.

2.2. TGF β signalling affects expression of bone morphogenetic protein antagonists and extracellular matrix remodelling

Many genes have been linked to coloboma [12,13]. However, optic fissure fusion is still not well understood on the structural and the molecular level. For optic fissure fusion to occur, the ECM has to be remodelled intensively. TGF β signalling is well known for its control of ECM remodelling in various processes [23,28,39]. We thus addressed the potential transcriptional ECM regulation during optic fissure fusion using our coloboma model. We quantified the levels of mRNAs from E13.5 embryonic eyes using Agilent microarrays. To this end we compared RNA harvested from eyes of wild-type embryos and TGF β 2/GDNF double mutant embryos, in which the coloboma phenotype was assigned to TGF β 2 function.

Notably, we found the expression of two BMP antagonists, follistatin (Fst) and gremlin (Grem)1, was downregulated in the coloboma model (figure 1e). We validated the regulation of these genes by quantitative PCR comparing RNA from eyes of TGF β ^{+/+} GDNF^{+/-} and TGF β 2^{-/-} GDNF^{+/-} embryos (figure 1f).

Furthermore, we processed the obtained microarray data, focusing on significantly downregulated genes. Performing bioinformatics analysis, we found as most prominent terms ECM, ECM organization, mesenchyme development,

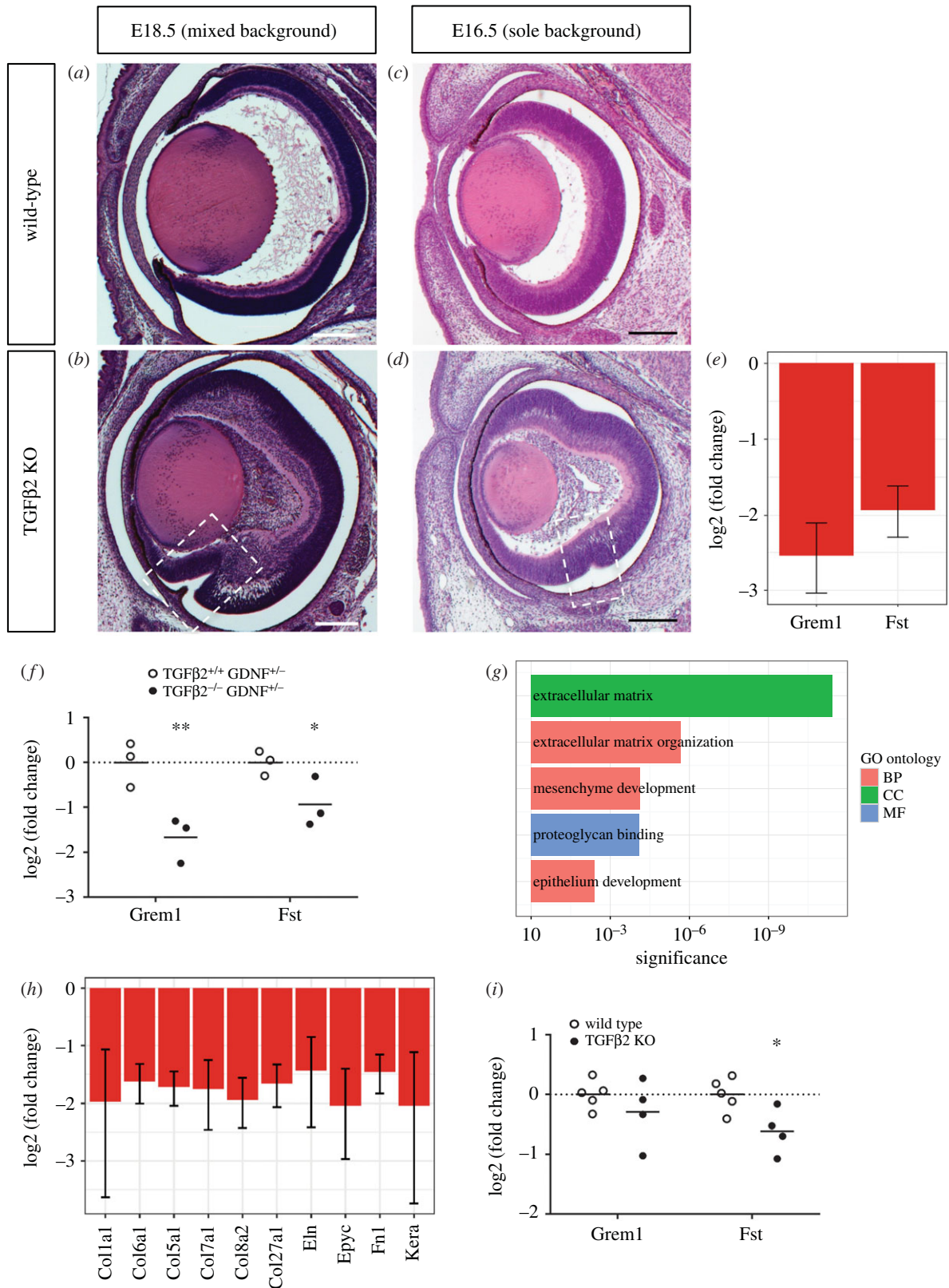


Figure 1. Loss of TGFβ2 ligand results in coloboma. (a,b) Frontal sections (E18.5, H&E) of (b) TGFβ2 KO and (a) wild-type mouse embryos from mixed genetic background. Note the persisting optic fissure (boxed in b), scale bars 200 μm. (c,d) Frontal sections (E16.5, H&E) of (d) TGFβ2 KO and (c) wild-type mouse embryos from a sole genetic background, mild coloboma phenotype boxed in (d), Scale bars 200 μm. (e) Expression analysis of gremlin and follistatin, decrease in TGFβ2 KO (TGFβ2/GDNF KO) as represented by the log₂(fold change). Error bars represent the 95% confidence interval. Corrected *p*-values of control gene expression compared to KO for Grem1 and Fst, 5.5×10^{-3} and 1.2×10^{-3} , respectively. (f) Expression analysis of gremlin and follistatin by quantitative PCR, decrease in TGFβ2 KO (TGFβ2^{-/-}, GDNF^{+/-}) compared to controls (TGFβ2^{+/+}, GDNF^{+/-}) as represented by the log₂(fold change) of individual samples; *n* = 3, horizontal bars represent the arithmetic mean. *p*-values for Grem1 and Fst, 7.8×10^{-3} and 0.029, respectively. (g) Selected terms enriched in the set of downregulated genes in TGFβ2/GDNF KO microarray based on gProfiler analysis. BP, biological process; CC, cellular component; MF, molecular function. (h) Expression analysis of several ECM-related genes. Error bars represent the 95% confidence interval. (i) Expression analysis of gremlin and follistatin by quantitative PCR, differential expression in TGFβ2 KO (sole genetic background) compared to wild-type as represented by the log₂(fold change) of individual samples; *n* = 5, one KO sample was excluded as an outlier. Horizontal bars represent the arithmetic mean. *p*-Values for Grem1 and Fst, 0.312 and 0.027, respectively.

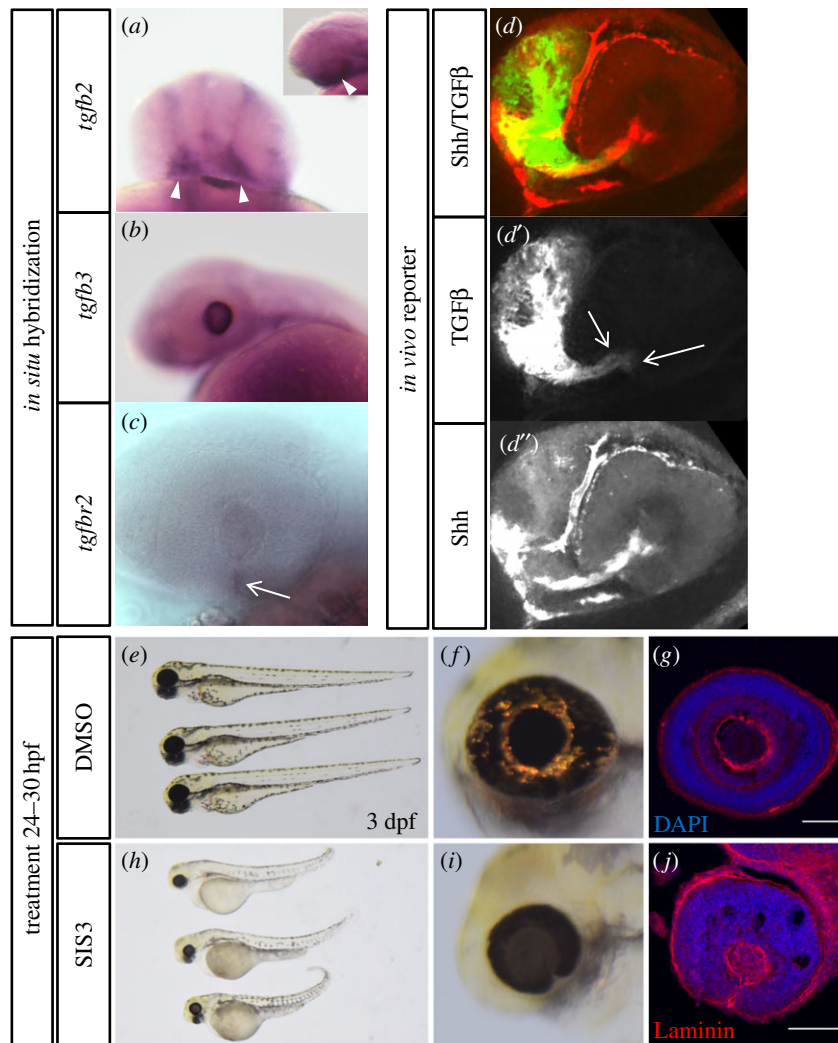


Figure 2. TGF β in the zebrafish eye. (a) Whole-mount *in situ* hybridization (WMISH) of *tgfb2* (30 hpf), frontal view. Small image shows a lateral view. Note expression in periocular tissue (arrowheads). (b) WMISH of *tgfb3* (30 hpf), lateral view. Note expression in the developing lens. (c) WMISH of *tgfb2* (30 hpf), lateral view. Note expression in the optic fissure (arrow). (d) *In vivo* signalling for TGF β (green) and Shh (red) for orientation, split into TGF β (d') and Shh (d''), at 24 hpf. Active TGF β signalling in the optic fissure margins (d', arrows). (e–j) Embryos (3 dpf) treated with DMSO (e–g) or specific inhibitor of Smad3 (SIS3, h–j) from 24 to 30 hpf. Sagittal sections through eyes of (g) DMSO and (j) SIS3-treated embryos at 3 dpf, stained with DAPI and anti-Laminin. SIS3-treated embryos show a persisting optic fissure with persisting basal lamina and absence of retinal layering.

epithelium development or proteoglycan binding (figure 1g). We found lower expression of collagen genes *Col1a1*, *Col6a1*, *Col5a1*, *Col7a1*, *Col8a2* and *Col27a1* in combination with lower levels of elastin (*Eln*) and epiphygan (*Epyc*) (figure 1h), pointing towards lower levels of fibrillogenesis in the extracellular space.

In addition, we isolated RNA from eyes dissected from TGF β 2 single mutants from a sole background at E12.5. The shift to E12.5 was necessary due to the fact that in the sole background condition the optic fissure fusion was occurring earlier than in the mixed background condition. Focusing on the interaction between TGF β and BMP signalling, we quantified the expression of the two BMP antagonists. We found a significant downregulation of *Fst*, but no significant downregulation of *Grem1* (figure 1i).

2.3. TGF β ligands, the ligand binding receptor and TGF β signalling in zebrafish optic cups

Next, we wanted to further address the functional role of TGF β signalling for optic fissure fusion and the interplay with BMP

signalling. To this end we switched to zebrafish (*D. rerio*). To ensure that a switch of model system to zebrafish is feasible, we investigated the expression of TGF β ligands and the TGF β ligand binding receptor during zebrafish eye development. We found *tgfb2* expressed in periocular tissue (figure 2a) whereas *tgfb3* was expressed in the developing lens (figure 2b). The ligand binding receptor *tgfb2b* we found expressed at the site of the optic fissure (figure 2c). To assess the dynamics of activated TGF β signalling *in vivo* during zebrafish development, we established a transcriptional TGF β sensor in a transgenic zebrafish line. The reporter system is based on Smads, the canonical transcription factors transducing TGF β signalling [40]. We used repetitive Smad binding elements (SBEs) from the human plasminogen activator inhibitor (PAI) (electronic supplementary material, figure S2A). Such a reporter has been intensively used for years as a luciferase assay to assess the amount and activity of TGF β in cell culture [41] and in mice [42]. We then established a transgenic zebrafish line. Activated TGF β signalling can be observed during development, e.g. in the forebrain region as well as in the distal tail (electronic supplementary material, figure S2B). We validated the functionality of this reporter line employing the

established TGF β signalling inhibitor SB431542 [43,44] (electronic supplementary material, figure S2C). Next, we wanted to know whether TGF β signalling was active in the optic fissure margins. We employed the TGF β signalling reporter line in combination with a reporter for Shh signalling (analogous to [45]) to relate the fissure to the optic stalk and performed imaging. We found the TGF β signalling reporter active within the optic fissure margins (figure 2*d,d'*, see Shh-reporter activity for orientation (*d''*)). Taken together, our data indicate that TGF β signalling is indeed active in the optic fissure margins of the zebrafish. Furthermore, we wanted to test if TGF β signalling is functionally involved in optic fissure fusion in zebrafish. In order to do this, we applied the compound inhibitor specific inhibitor of Smad3 (SIS3) to wild-type embryos during the onset of optic fissure fusion from 24 to 30 hpf. SIS3 prevents phosphorylation of Smad3 [46], which, together with Smad2, is the main transcription factor in the canonical TGF β signalling pathway. Treatment with SIS3 yielded smaller embryos at 3 dpf (figure 2*h,i*; see *e,f* as control). The tail extension as well as the brain development seemed affected, matching the expression domains we observed in the TGF β reporter embryos (compare electronic supplementary material, figure S1B, C). Most importantly, the treated embryos had coloboma with a persisting basal lamina at 3 dpf in conjunction with reduced eye size (figure 2*i,j*).

2.4. TGF β mediated bone morphogenetic protein antagonism during optic fissure closure

We found two antagonists for BMP signalling, follistatin and gremlin1, transcriptionally downregulated in our murine coloboma model (figure 1*e*). BMP4 in combination with Vax2 is important for the definition of cellular identities along the dorsal ventral axis within the vertebrate eye [47–50]. In line with these data we found *bmp4* expressed dorsally and *vax2* expressed ventrally within the zebrafish optic cup (figure 3*a,b*). Furthermore, we found homologous genes to the identified BMP antagonists (*grem2b* and *fsta*) expressed in the optic fissure margins of zebrafish also opposing the *bmp4* expression domain (figure 3*c–f*). The expression of *grem2b* also matched the expression pattern previously reported for the gene [51]. We also tested whether these antagonists are regulated by TGF β in fish by comparing their expression in SIS3-treated embryos at 30 hpf with DMSO-treated controls using quantitative PCR. Inhibition of Smad3 signalling caused downregulation of *grem2b*, but not *fsta* (figure 3*g*). However, because we extracted RNA from embryonic heads and *fsta* is also expressed in the ciliary marginal zone and some parts of the brain, it would be plausible that a possible regulation by TGF β exclusively in the optic fissure would not be detectable. Alternatively, it is also possible that *fsta* is not regulated by Smad3 or TGF β at all in zebrafish.

BMP4 is a secreted ligand and can potentially diffuse and act over extended distances. The expression of the two BMP antagonists in the optic fissure hints at a functional requirement to locally suppress BMP activity in this domain. Our data indicate that TGF β signalling is relevant for ECM remodelling during optic fissure fusion. Notably, BMP signalling was shown to potentially counteract such TGF β -induced changes [29–32]. We propose that TGF β -induced local BMP antagonism is protecting the expression of TGF β -regulated genes, which are facilitating ECM remodelling during optic fissure

fusion (figure 3*h*, scheme). We next wanted to functionally test our hypothesis. We generated a transgenic line allowing heat shock inducible *bmp4* expression (*tg(hsp:bmp4 cmlc2:GFP)*) (figure 3*i*). With this heat shock inducible transgenic line, we aimed at an oversaturation of the BMP antagonists within the optic fissure margins. Since the morphogenesis of the optic cup is dependent on BMP antagonism [11], the timing of the heat shock induced expression of *bmp4* was critical. Thus, we tested the outcome of the heat shock induced *bmp4* expression at different successive stages of development (figure 3*i*). Induced expression at 21 hpf and 22 hpf resulted in an extended coloboma (figure 4*a–a''* and *b–b''*, see *d–e''* as control), well in line with the coloboma observed in our previous analyses [11]. This indicates that the transgenic line is functional and sufficiently high BMP4 levels are expressed but it also indicates that the onset of induction was too early and was affecting optic cup morphogenesis. The induced expression of *bmp4* at 23 hpf resulted in a milder coloboma, with less affected cup morphogenesis (figure 4*c–c''*). Notably, the coloboma phenotypes resulting from *bmp4* expression induced at 24, 25 and 26 hpf were comparable. Importantly, they were not showing defects in optic cup morphogenesis (figure 4*f–h''*, see *i–j''* as control, and figure 4*k–n*). In the proximal part of the optic cup, the optic fissure margins were closely aligned but not fused (figure 4*k'*). Thus, we identified this phenotype as a defect in optic fissure fusion.

Next, we wanted to know whether the heat shock induced overexpression of *bmp4* affected the downstream signalling targets of TGF β in the optic fissure. Therefore, we extracted RNA for quantitative analysis from the heads of 30 hpf *tg(hsp:bmp4 cmlc2:GFP)* embryos which had been subjected to a heat shock at 24 hpf. Wild-type siblings from the same clutch of eggs served as a control. We found that *grem2b* was downregulated in the *bmp4* overexpression condition by trend, although this result was not statistically significant ($p = 0.17$), while *fsta* was strongly upregulated (figure 5*a*). This upregulation was also seen in *in situ* hybridization of *fsta* and affected the expression domains in the trunk, the brain and the optic stalk and optic fissure, but not the ciliary marginal zone (figure 5*b,c*).

3. Discussion

Many genes and signalling pathways have been linked to coloboma [2–11] and a resulting coloboma gene network [12,13] is growing. The morphology of the coloboma phenotypes, however, is highly variable and many phenotypes likely result from early morphogenetic defects (e.g. [11]). On the other hand, little still is known about the process of optic fissure fusion. Recently, it was shown that hyaloid vessels are important for basement membrane degradation [22], a process important at the initiation of optic fissure fusion. This process is likely involving the transcription factors Pitx2 [52], Pax2 [14] and Vax2 [53] and potentially also retinoic acid signalling [52]. However, with respect to the subsequent process, the actual fusion of the optic fissure margins, less data is available.

It is unclear how the structure of the epithelial margins is loosened and eventually disassembled locally and how new connections are established, linking the neuroretinal domains and the RPE domains of both margins. Especially for the epithelial disassembly, the molecular mechanism is elusive. Concerning the formation of new connections, *n-cadherin* and

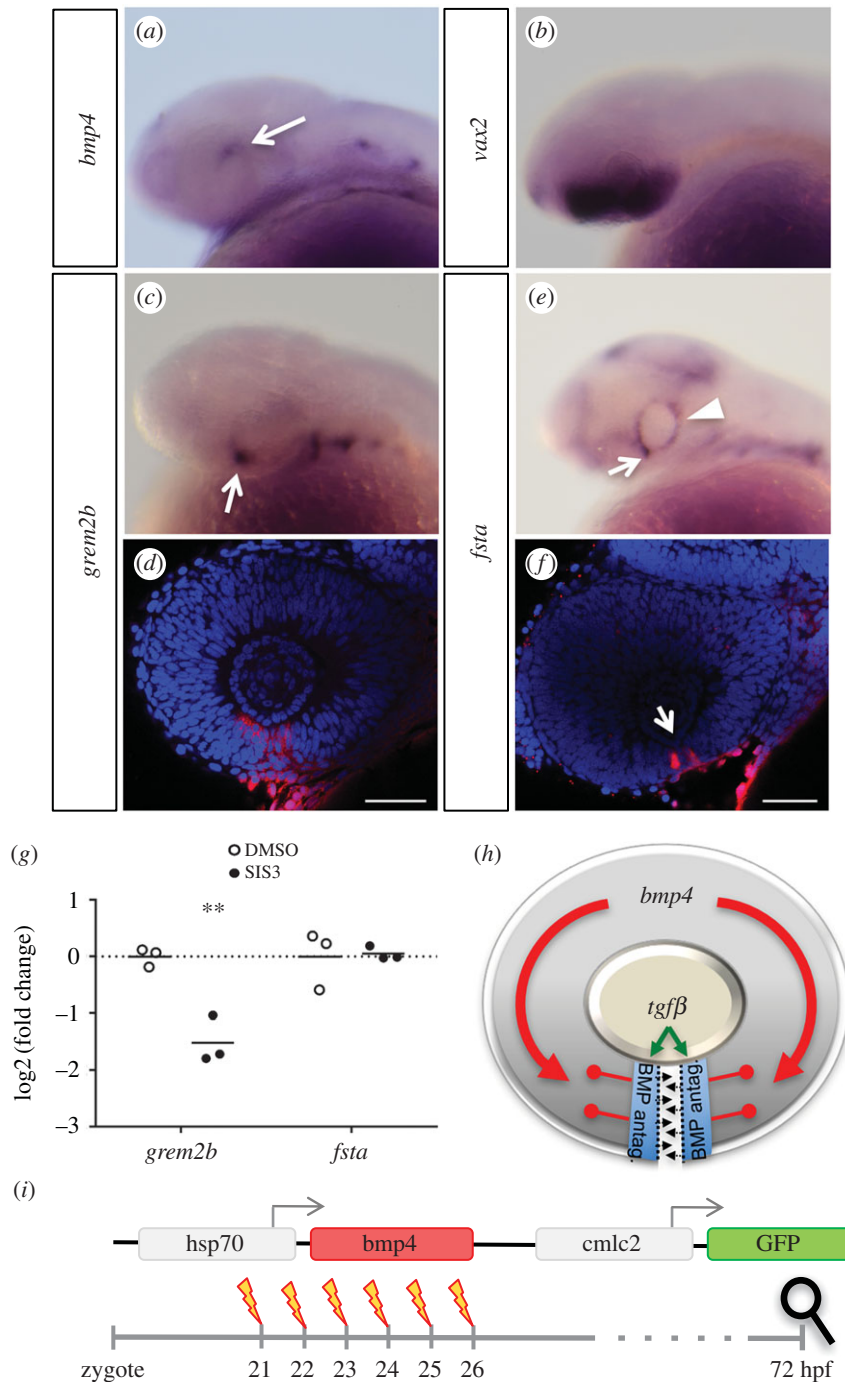


Figure 3. BMP antagonists *grem2b* and *fsta* are expressed in the optic fissure. WMISHs were performed at 30 hpf and are shown in lateral view. (a) *bmp4* is expressed in the dorsal optic cup (arrow). (b) Expression of *vax2* in the ventral retina. (c,d) Expression of *grem2b* in the optic cup is restricted to the optic fissure (arrow). (d) Confocal view of *grem2b* expression (red) with DAPI counterstaining (blue). (e,f) *fsta* is expressed in the optic fissure (arrows), as well as the ciliary marginal zone (CMZ, arrowhead). (f) Confocal view of *fsta* expression (red) with DAPI counterstaining (blue). (g) Expression analysis of gremlin and follistatin by quantitative PCR, differential expression in heads of SIS3-treated embryos (30 hpf) as represented by the log₂(fold change) of individual samples. Embryos were treated from 24 hpf onward, controls were treated with DMSO. Material from three individuals was pooled for one sample; $n = 3$, horizontal bars represent the arithmetic mean. p -Values for *grem2b* and *fsta*, 4.3×10^{-3} and 0.877, respectively. (h) Model of the proposed role of TGFβ and BMP antagonism during optic fissure fusion. TGFβ signalling domains in the optic fissure margins are shielded from BMP by induced BMP antagonists. (i) Scheme of a heat shock inducible BMP construct used to create the transgenic line *tg(hsp70:bmp4, cmcl2:GFP)*. GFP expressed under the cardiac *cmcl2* promoter serves as transgenesis marker. Experimental procedure using heat shocks at different time points between 21 and 26 hpf to induce *bmp4* expression, aiming at disrupting optic fissure fusion. Analysis of phenotypes was scheduled for 3 dpf.

α -catenin [20,21] are good candidate genes. The morphology of the coloboma resulting from a conditional α -catenin mutant shows an alignment of the margins [21], indeed indicating a functional involvement related to the fusion process. The coloboma resulting from *n-cadherin* mutation, however, shows a certain remaining gap [20], suggesting that not the process of fusion but a preceding event was affected by the mutation.

Our data indicate that TGFβ signalling is necessary for the fusion of the optic fissure margins. Hereby, we are integrating TGFβ as a new member into the coloboma gene network. Besides, TGFβ signalling is also known to be essential for palatal fusion [33,54]. Notably, the ligands TGFβ2 and TGFβ3 have slightly different functions there. While in TGFβ2 mutant mice the palatal shelves stay apart and a gap remains [33], in TGFβ3

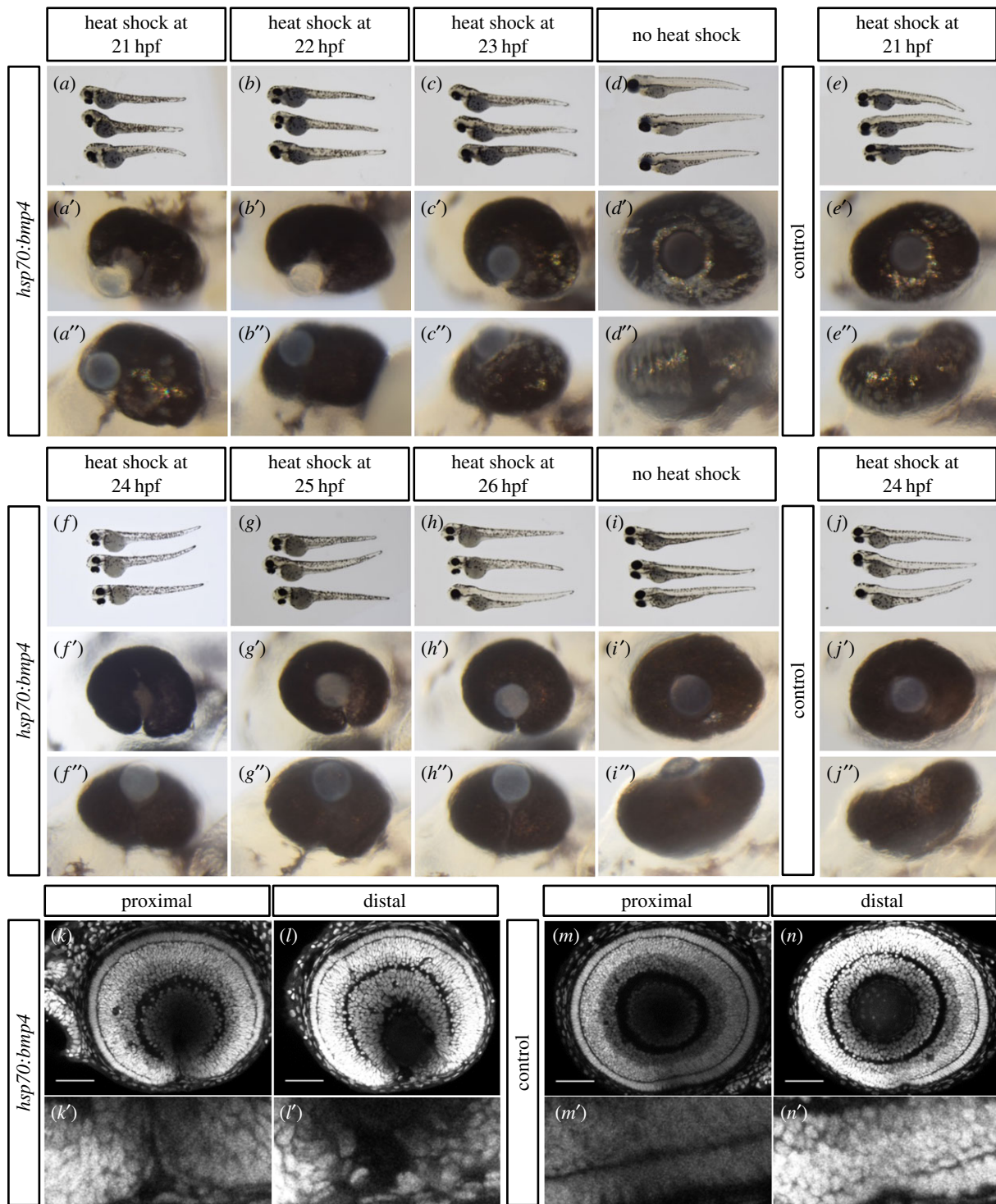


Figure 4. Differentially timed induction of *bmp4* causes different coloboma phenotypes. (a–e) Gross morphology of (a–d) *tg(hsp70:bmp4, cmlc2:GFP)* embryos after heat shock at 21, 22, 23 hpf, no heat shock, and (e) wild-type embryos heat shocked at 21 hpf. (a'–h') Close up, lateral view, (a''–h'') close up, ventral views. Early *bmp4* induction impairs optic cup morphogenesis, resulting in coloboma (a'–c''). (f–j) Gross morphology of (f–i) *tg(hsp70:bmp4, cmlc2:GFP)* embryos after heat shock at 24, 25, 26 hpf, no heat shock and (j) wild-type embryos heat shocked at 24 hpf. (f'–j'') Close up, lateral view, (f''–j'') close up, ventral views. Late *bmp4* expression hampers optic fissure fusion, resulting in coloboma (f'–h''). All animals were visualized at 3 to 3.5 dpf. (k–n) Lateral confocal images of larval eyes (3 dpf) stained with DAPI. Individuals were heat shocked at 24 hpf. *Tg(hsp70:bmp4, cmlc2:GFP)* animals show a persisting optic fissure (k,l) while their wild-type siblings do not (m,n). (k'–n') are magnifications of the optic fissure region. Notably, the fissure margins are touching in the proximal part of the eye (k').

mutants the palatal shelves potentially touch, but do not fuse [54,55]. The latter is reminiscent of what we observed in our study: the fissure margins meet, but do not fuse (figure 1b).

During our analysis we observed a potential influence of the genetic breeding background on optic fissure closure. Firstly, optic fissure fusion seemed to occur one day earlier in wild-type embryos from the sole breeding background

(around E13) compared to those from the mixed breeding background (around E14). Secondly, the marked, strong coloboma phenotypes can be observed in both $TGF\beta 2^{-/-}$ $GDNF^{-/-}$ and $TGF\beta 2^{-/-}$ $GDNF^{+/+}$ embryos from a mixed breeding background, occasionally also presenting a dorsal coloboma, while the phenotype of $TGF\beta 2$ KO embryos from a sole breeding background is noticeably milder (figure 1a–d, electronic

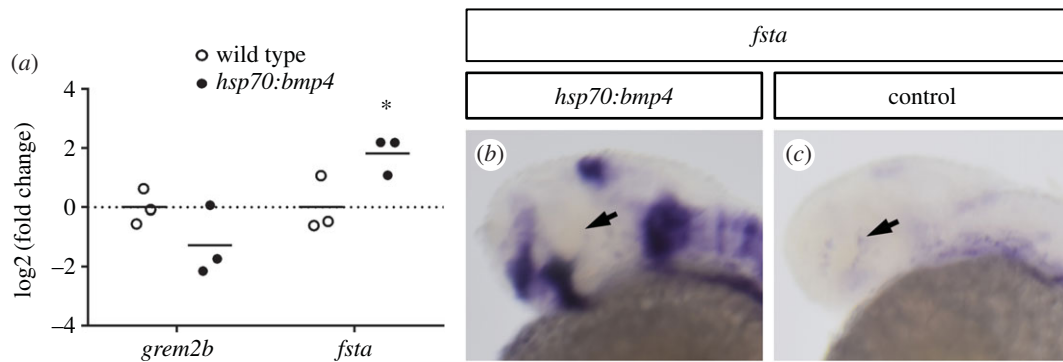


Figure 5. Timed induction of *bmp4* affects expression of BMP antagonists. (a) Expression analysis of gremlin and follistatin by quantitative PCR, differential expression in heads of *tg(hsp70:bmp4, cmlc2:GFP)* embryos at 30 hpf after heat shock at 24 hpf as represented by the log₂(fold change) of individual samples. Material from three individuals was pooled for one sample; $n = 3$, horizontal bars represent the arithmetic mean. p -Values for *gremlin* and *fsta*, 0.170 and 0.049, respectively. (b,c) WISH for *fsta* (30 hpf) in *tg(hsp70:bmp4, cmlc2:GFP)* after heat shock at 24 hpf (b) and control embryos (c). Strong upregulation after the heat shock is seen in the optic fissure and other expression domains, but not the ciliary marginal zone (arrow). WISHs in (b) and (c) were stained in parallel for the same amount of time.

supplementary material, S1B–C). Lastly, in gene expression analysis, we found significant downregulation of *Grem1* and *Fst* in $TGF\beta 2^{-/-}$ $GDNF^{+/-}$ eyes compared to $TGF\beta 2^{+/+}$ $GDNF^{+/-}$ eyes (figure 1*f*), whereas in the $TGF\beta 2$ KO derived from a sole breeding background, *Grem1* was not significantly regulated and *Fst* downregulation was less pronounced (figure 1*i*). We want to mention that these effects could also be due to or maybe just influenced by a yet unreported sensitizing effect of GDNF signalling. Even if GDNF single KO embryos do not show a coloboma and the transcriptional analyses performed during our analyses contradict, we cannot totally rule out that an affected GDNF signalling sensitizes the system to the loss of $TGF\beta 2$. An interaction between these two $TGF\beta$ subfamilies ($TGF\beta$ and GDNF) is well known [56–58]. However, the signalling interaction in such scenarios is occurring vice versa, and $TGF\beta$ was found to facilitate GDNF signalling. Overall it seems more likely that the documented ocular GDNF expression [38] is important for later stages of development, because GDNF was previously reported to promote the proliferation and differentiation of photoreceptors during eye development [59]. It is expressed in the lens and retina of E14.5 mouse embryos [38], which is after the closure of the optic fissure, during differentiation of the neuroretina. Taken together, we consider it more likely that the differences we observe in phenotypes and transcriptomes are due to the genetic background of the mouse strains used, although we cannot rule out a contribution of GDNF based on our data.

$TGF\beta$ signalling is well-described to be an important regulator of modifications to the ECM in development and disease [23–28]. $TGF\beta$ -induced changes to the ECM are, however, potentially counteracted by BMP signalling [29–32]. BMP signalling and BMP antagonism are crucial during many important steps of eye development, such as optic fissure generation [60], dorsal–ventral cell specification and optic cup formation [11]. In the latter study, we showed that an inhibition of BMP signalling is crucial for a bilateral neuroretinal flow to be maintained over the distal rim of the developing optic cup. This BMP signalling inhibition was achieved by the BMP antagonist follistatin (*fsta*). Considering also the early expression domains of *gremlin* [51] it seems likely that these two BMP antagonists also cooperate during optic cup formation during neuroretinal flow maintenance.

Here, we propose that $TGF\beta$ -induced local BMP antagonism in the optic fissure is protecting the expression of $TGF\beta$ -regulated genes which facilitate fissure fusion. A similar process, involving the BMP antagonist gremlin, was observed in the context of glaucoma [61]. Gremlin has, furthermore, been linked to cleft lips in humans [62], indicating that the level of BMP signalling must be tightly controlled during fusion processes there as well. A locally induced BMP antagonism within the fissure margins seems especially important because also other BMP ligands besides *bmp4* are expressed within the optic cup, e.g. *bmp2b*, *bmp2a* and *bmp6* and *bmp7b* [63–66]. The combined expression of two BMP antagonists within the optic fissure margins is likely important to provide robustness to the system. BMP antagonists are often expressed redundantly [67–69], pointing at the importance of their function. Thus, it is not surprising that the loss of a single BMP antagonist does not result in coloboma (*Grem1* mutant mice). In $TGF\beta 2$ KO embryos with a sole breeding background, we found that *Grem1* was noticeably less downregulated compared to embryos with a mixed breeding background, while *Fst* downregulation was changed only slightly. This might explain the less severe phenotype that we observed in the colobomatous embryos from a sole breeding background (figure 1*d*).

While our data strongly indicate that BMP antagonism is important for optic fissure fusion, we do not yet understand how BMP signalling can interfere with this process, specifically how it could counteract the changes induced by $TGF\beta$ signalling. BMP might repress the expression of $TGF\beta$ receptors or signal transduction components directly. In mouse pulmonary fibroblasts, BMP was shown to reduce $TGF\beta$ -dependent collagen expression by inducing inhibitors of differentiation 2 and 3 (*Id2/3*) [29]. The same study reported that nuclear localization of Smad3 in response to $TGF\beta$ signalling was inhibited by BMP, while another study found that Smad1 and Smad2/3 co-localized in nuclei of renal tubules in response to simultaneous $TGF\beta$ and BMP signalling [30]. The inhibitory Smad7 [70] could potentially also be involved in the signalling interaction. In mouse, knockout of the Smad7 gene led to various eye defects including coloboma [71]. Thus, it appears that there is a spectrum of $TGF\beta$ –BMP interaction which depends on the biological context and may include transcription-dependent and -independent mechanisms. Further research is needed to determine the mode of interaction between the

two pathways in the context of tissue fusion and whether BMP signalling is directly inhibiting the TGF β signalling pathway or only downstream targets of TGF β . Our results suggest that only a subset of TGF β target genes might be inversely regulated by BMP signalling but that these genes would be crucial for the optic fissure fusion process.

Our data obtained by timed overexpression of *bmp4* in zebrafish embryos clearly indicate that BMP induction has a detrimental effect on optic fissure fusion, resulting in coloboma (figure 4). We showed, in accordance with our previous findings [11], that *bmp4* overexpression during optic cup formation (prior to 24 hpf) causes morphogenetic defects by disrupting epithelial flow. Only overexpression after 24 hpf was able to induce a defect of optic fissure fusion. Under this condition, we also observed a repression of the TGF β target gene *grem2b* by trend (figure 5a). However, this result was not statistically significant, thus leaving the relevance unclear. By contrast and unexpectedly, *fsta* was found upregulated after forced *bmp4* expression (figure 5). This could be a response of the developing organism to the increased levels of BMP, trying to reduce BMP signalling to a physiological level by upregulating *fsta*. Notably, the fissure margins were responding intensively in comparison to the ciliary marginal zone, where hardly any change of expression was noticeable. Furthermore, *fsta* was not found downregulated in response to SIS3 treatment (figure 3g). This could imply that *fsta* is not regulated by Smad3, at least in zebrafish. It is however still possible that it is instead regulated through Smad2. Several genes are known that possess binding sites for only one of these two Smads, including Goosecoid (*gsc*) and *mix2* for Smad2, as well as JunB/*junba* and PAI-1/*serpine* for Smad3 [41,72–74]. Another interpretation would be that *fsta* is not regulated by TGF β in zebrafish.

It should be mentioned that a completely different interpretation is also supported by our data, in which the upregulation of *fsta* is not a compensatory effect but the cause for the optic fissure closure defect that we observe after BMP overexpression. In that case, the mechanism would be independent of TGF β . While we cannot exclude this option, we consider it unlikely because *fsta* is downregulated in the TGF β 2 KO mouse model. In summary, we conclude that *fsta* is either not controlled by Smad3 or TGF β in zebrafish, or that due to differential regulation of the expression domains in the fissure margins and the ciliary marginal zone, a subtle regulation within only the margin domain was not detectable by qPCR.

4. Conclusion

The process of optic fissure fusion is not well understood, especially not on a cellular and molecular basis. We found an interplay of two growth factors, TGF β and BMP, during the fusion of the optic fissure margins. While TGF β signalling is acting pro-fusion and induces changes to the ECM, we found that BMP signalling is capable of inhibiting fissure fusion. Notably, TGF β signalling is inducing BMP antagonists within the fissure margins. This finding suggests that thereby TGF β , itself acting pro-fusion, is also locally counteracting BMP signalling, which is acting anti-fusion. Even if gene regulation might differ depending on the breeding background and between fish and mouse, the functional concept appears to be conserved. Our findings can likely be applied also to other fusion processes, especially when TGF β signalling or BMP antagonism is involved, as in fusion processes during orofacial

development. Together with our previous data, our current work indicates a dual role of BMP antagonism, first during optic cup formation, maintaining a bilateral neuroretinal flow entering the optic cup, and second during optic fissure fusion.

5. Material and methods

5.1. Mice

For this study TGF β 2^{+/-} [33] and GDNF^{+/-} [35] mice were used for breeding. Timed matings were performed overnight and the day on which a vaginal plug was visible in the morning was considered as day 0.5. Analyses were restricted to embryonic stages because of perinatal lethality of the individual mutants. For analysis of embryonic tissue, the mother was sacrificed and the embryos were collected by caesarean section. All of the experiments were performed in agreement with the ethical committees. Genotyping was performed according to Rahhal *et al.* [34]. The term ‘TGF β 2 KO with mixed breeding background’ refers to a TGF β 2^{-/-}, GDNF^{+/+} offspring of TGF β 2^{+/-}, GDNF^{+/-} parents, because the original single mutant lines were created in different wild-type strains [33,35]. A ‘sole breeding background’ refers to the TGF β 2 line [33]. This line was maintained and crossed with C57BL/6.

5.2. Histological analysis

Tissue was processed for paraffin sectioning. Frontal sections of control and TGF β 2^{-/-} embryos were obtained and stained with haematoxylin and eosin.

5.3. Microarray data

RNA was extracted from whole eyes of E13.5 embryos (controls and TGF β 2^{-/-} (GDNF^{-/-}) respectively). RNA was reverse transcribed, amplified and loaded on Agilent one-colour microarray chips. Experiments were performed in triplicates.

Further analysis was performed using R [75] and the bioconductor packages Agi4 \times 44PreProcess, limma and mgug4122a.db as annotation database. For background correction we used the following parameters: BGmethod = ‘half’, NORM-method = ‘quantile’, foreground = ‘MeanSignal’, background = ‘BGMedianSignal’ and offset = 50. The probes were filtered using the recommended thresholds and afterwards the replicated non-control probes were summarized. Then the method *lmFit* was used to fit a linear model on the arrays. Finally, the differential expression statistics were computed using the methods *eBayes*.

Next only those genes with fold change higher than 1.5 were considered, then a multiple comparison correction was performed on the *p*-values using the BH (Benjamini and Hochberg) method. The genes with corrected *p*-value lower than 0.05 were defined as significantly differentially expressed genes. The microarray data supporting this article have been made available via the repository ‘BioStudies’ under the identifier S-BSST80.

5.4. Functional analysis of gene sets

We used the tool gProfiler ([76], <http://biit.cs.ut.ee/gprofiler/>) version 6.7 to find enriched terms on the set of significantly downregulated genes from the mouse arrays. We provided the official gene symbol of these genes as input and used the default set of databases.

5.5. RNA extraction from mouse and zebrafish tissue for quantitative PCR

RNA was extracted from whole eyes of E12.5 embryos (controls and $TGF\beta 2^{-/-}$, respectively). For zebrafish, three heads from 30 hpf embryos were pooled for each sample. RNA was extracted using the guanidinium thiocyanate–phenol–chloroform method (modified after [77]). After DNaseI treatment, it was purified again by phenol–chloroform extraction and sodium acetate precipitation.

5.6. Quantitative PCR

RNA was reverse transcribed with the ProtoScript II First Strand cDNA Kit (New England Biolabs). Quantitative real-time PCR was performed with a CFX Connect Real-Time PCR Detection system (Bio-Rad Laboratories) and the Luna Universal qPCR Master Mix (New England Biolabs) in technical triplicates with 20 μ l reaction volume and 2 μ l of a 1:10 dilution of the cDNA template. *Gapdh* was used as reference gene for mouse qPCR and *efl1a11* was used for zebrafish qPCR. For primers used, see electronic supplementary material Information.

For data analysis, the dCq values were calculated by subtracting $Cq_{[target\ gene]}$ from $Cq_{[reference\ gene]}$, and ddCq values were calculated by subtracting the averaged $dCq_{[control]}$ from the simple $dCq_{[treatment/KO]}$ [78]. These ddCq values are presented as $\log_2(\text{fold change})$. *p*-Values were determined by two-tailed, unpaired *t*-test, except for the data presented in figure 1f. There we used a one-tailed test because we knew to expect downregulation from the microarray dataset.

In figure 5a, the expression data for *grem2b* derives from two runs with the same samples (biological replicates) in the same qPCR cyclers. For this, an average dCq value was calculated for each biological replicate.

5.7. Zebrafish husbandry

Zebrafish (*D. rerio*) were kept as closed stocks in accordance with local animal welfare law and with the permit 35-9185.64/1.1 from the Regierungspräsidium Freiburg. The fish were maintained in a constant recirculating system at 28°C on a 12 L : 12 D cycle. Fish lines used in this study were created in the AB wild-type strain.

5.8. Transgenic zebrafish

Plasmid DNA containing SBEs in combination with a minimal promoter were kindly provided by Peter tenDijke ((CAGA)₁₂ MLP Luc). Here, repetitive SBEs derived from the promoter of the human PAI gene [41] were used to drive a luciferase gene.

We cloned the SBEs with the minimal promoter into a Gateway 5' entry vector (Invitrogen). A multisite Gateway reaction [79] was subsequently performed resulting in an SBE driven GFPcaax construct (SBE:GFPcaax). A zebrafish line was generated with SB (sleeping beauty) transgenesis according to Kirchmaier *et al.* [80]. Shh reporter zebrafish were generated according to Schwend *et al.* [45]. The plasmid was kindly provided by Sara Ahlgren.

We assembled the expression construct for *Tg(hsp70:bmp4, cmlc2:eGFP)* in a Gateway reaction, using a Tol2 destination vector including *cmlc2:eGFP* [79], a 5' entry vector containing

the *hsp70* promoter, a pENTR D-TOPO (ThermoFisher Scientific) vector containing the CDS of *bmp4* [11] and a 3' entry vector with a polyadenylation site [79]. The construct (10 ng μ l⁻¹) was injected into wild-type zebrafish zygotes together with Tol2 transposase mRNA (7 ng μ l⁻¹) [79]. Embryos with strong GFP expression in the heart were selected as founders. Lines were kept in closed stocks and validated in every generation.

5.9. Drug treatments

Zebrafish embryos were treated with SIS3 (9 μ M, 3 mM stock in DMSO [81]) and with SB431542 (80 μ M, 10 mM stock in DMSO [82]). Controls were treated with DMSO without the inhibitor.

5.10. Microscopy

Signalling reporter fish were imaged with a Leica SP5 setup, samples were mounted in glass bottom dishes (MaTek). For time-lapse imaging embryos were embedded in 1% low melting agarose covered with zebrafish medium and anaesthetized with tricaine. Left and right eyes were used and oriented to fit the standard views. A stereomicroscope (Olympus/Nikon) was used for recording bright field and fluorescent images of TGF β signalling reporter fish. Whole-mount *in situ* hybridizations were recorded with a stereomicroscope (Nikon SMZ18) as well as an upright microscope (Zeiss) and a confocal Leica SP8 setup.

5.11. Heat shocks and controls

tg(hsp70:bmp4, cmlc2:eGFP) eggs were kept in a Petri dish at 28°C after fertilization. To induce *bmp4* expression, 21–26 hpf embryos were transferred to a 1.5 ml reaction tube and incubated for 1 h at 37°C in a heating block. Afterwards, they were returned to a dish at 28°C. Embryos were fixed with 4% PFA at 30 hpf for *in situ* hybridization and at 3 dpf for morphological analysis.

We used *tg(hsp70:bmp4, cmlc2:eGFP)* embryos which were not heat shocked as controls, as well as heat shocked wild-type siblings from the same clutch of eggs.

5.12. Whole-mount *in situ* hybridization

Whole-mount *in situ* hybridizations (WMISHs) were performed according to Quiring *et al.* [83]. WMISHs for confocal imaging were stained with FastRed Naphthol (Sigma-Aldrich).

5.13. Immunohistochemistry and confocal imaging of zebrafish embryos

Fixed embryos were bleached using 3% hydrogen peroxide and 0.5% potassium hydroxide. For whole-mount imaging, they were stained with 4 μ g ml⁻¹ 4',6-diamidino-2-phenylindole (DAPI) for 2 h. Imaging was performed in glass bottom dishes (Matek) using a Leica SP8 TCS setup.

For anti-Laminin staining, embryos were cryosectioned (20 μ m) and stained on Superfrost plus slides. Antibodies used: rabbit Laminin Ab-1 (ThermoFisher Scientific, RB-082-A1, 1:100), goat anti-rabbit-Alexa 555 (ThermoFisher Scientific, A-21428, 1:250).

Data accessibility. The datasets supporting this article have been uploaded as part of the supplementary material. The microarray data supporting this article have been made available via the repository 'BioStudies' under the identifier S-BSST80.

Authors' contributions. M.K. contributed to experiments and wrote the manuscript; J.M. performed analysis of microarray data; PE contributed to experiments; E.R. and B.R. contributed to TGF β KO mouse acquisition; A.Z. sent fixed embryonic heads (Grem1 KO and controls) for analyses; K.K. and J.W. helped conceive the study and acquired funding, S.H. conceived the study and experiments and wrote the manuscript.

Competing interests. The authors declare that they do not have a conflict of interest.

Funding. The project was in part funded by the Deutsche Forschungsgemeinschaft (Wi 1824/5-1, Kr 1477/14-1). The article processing

charge was funded by the German Research Foundation (DFG) and the Albert-Ludwigs-University Freiburg in the funding programme Open Access Publishing.

Acknowledgements. We want to thank Rolf Zeller for making gremlin KO mice available as well as for scientific discussion. We also want to thank Gabriela Salinas-Riester and the TAL of the University Göttingen. We also thank Sara Ahlgren for generously making the shh-reporter construct available and Peter tenDijke for the luciferase construct on which our new TGF β reporter zebrafish is based. We thank Lea Schertel, Ute Baur and Lidia Koschny for great technical assistance and Burkhard Höckendorf for constructive input and help generating the TGF β reporter line, and Hannes Voßfeld for assistance with the mouse mutants. We also thank Shokoufeh Khakipoor for helping to acquire mutant mouse embryos and Melanie Feuerstein for excellent technical assistance regarding qPCR.

References

- Onwochei BC, Simon JW, Bateman JB, Couture KC, Mir E. 2000 Ocular colobomata. *Surv. Ophthalmol.* **45**, 175–194. (doi:10.1016/S0039-6257(00)00151-X)
- Graw J. 2003 The genetic and molecular basis of congenital eye defects. *Nat. Rev. Genet.* **4**, 876–888. (doi:10.1038/nrg1202)
- Westenskow P, Piccolo S, Fuhrmann S. 2009 Beta-catenin controls differentiation of the retinal pigment epithelium in the mouse optic cup by regulating *Mitf* and *Otx2* expression. *Development* **136**, 2505–2510. (doi:10.1242/dev.032136)
- Bankhead EJ, Colasanto MP, Dyorich KM, Jamrich M, Murtaugh LC, Fuhrmann S. 2015 Multiple requirements of the focal dermal hypoplasia gene porcupine during ocular morphogenesis. *Am. J. Pathol.* **185**, 197–213. (doi:10.1016/j.ajpath.2014.09.002)
- Chen S *et al.* 2012 Defective FGF signaling causes coloboma formation and disrupts retinal neurogenesis. *Cell Res.* **23**, 254–273. (doi:10.1038/cr.2012.150)
- Cai Z, Tao C, Li H, Ladher R, Gotoh N, Feng G-S, Wang F, Zhang X. 2013 Deficient FGF signaling causes optic nerve dysgenesis and ocular coloboma. *Development* **140**, 2711–2723. (doi:10.1242/dev.089987)
- Miesfeld JB, Gestri G, Clark BS, Flinn MA, Poole RJ, Bader JR, Besharse JC, Wilson SW, Link BA. 2015 *Yap* and *Taz* regulate retinal pigment epithelial cell fate. *Development* **142**, 3021–3032. (doi:10.1242/dev.119008)
- Matt N, Ghyselinck NB, Pellerin I, Dupé V. 2008 Impairing retinoic acid signalling in the neural crest cells is sufficient to alter entire eye morphogenesis. *Dev. Biol.* **320**, 140–148. (doi:10.1016/j.ydbio.2008.04.039)
- Lupo G, Gestri G, O'Brien M, Denton RM, Chandraratna RAS, Ley SV, Harris WA, Wilson SW. 2011 Retinoic acid receptor signaling regulates choroid fissure closure through independent mechanisms in the ventral optic cup and periocular mesenchyme. *Proc. Natl Acad. Sci. USA* **108**, 8698–8703. (doi:10.1073/pnas.1103802108)
- Lee J, Willer JR, Willer GB, Smith K, Gregg RG, Gross JM. 2008 Zebrafish blowout provides genetic evidence for *Patched1* mediated negative regulation of Hedgehog signaling within the proximal optic vesicle of the vertebrate eye. *Dev. Biol.* **319**, 10–22. (doi:10.1016/j.ydbio.2008.03.035)
- Heermann S, Schütz L, Lemke S, Kriegstein K, Wittbrodt J. 2015 Eye morphogenesis driven by epithelial flow into the optic cup facilitated by modulation of bone morphogenetic protein. *Elife* **4**, e05216. (doi:10.7554/eLife.05216)
- Gregory-Evans CY, Williams MJ, Halford S, Gregory-Evans K. 2004 Ocular coloboma: a reassessment in the age of molecular neuroscience. *J. Med. Genet.* **41**, 881–891. (doi:10.1136/jmg.2004.025494)
- Gregory-Evans CY, Wallace VA, Gregory-Evans K. 2013 Gene networks: dissecting pathways in retinal development and disease. *Prog. Retin. Eye Res.* **33**, 40–66. (doi:10.1016/j.preteyeres.2012.10.003)
- Torres M, Gómez-Pardo E, Gruss P. 1996 *Pax2* contributes to inner ear patterning and optic nerve trajectory. *Development* **122**, 3381–3391.
- Favor J *et al.* 1996 The mouse *Pax2*(1 Neu) mutation is identical to a human *PAX2* mutation in a family with renal-coloboma syndrome and results in developmental defects of the brain, ear, eye, and kidney. *Proc. Natl Acad. Sci. USA* **93**, 13 870–13 875. (doi:10.1073/pnas.93.24.13870)
- Sanlaville D, Verloes A. 2007 CHARGE syndrome: an update. *Eur. J. Hum. Genet.* **15**, 389–399. (doi:10.1038/sj.ejhg.5201778)
- Fletcher J, Hu M, Berman Y, Collins F, Grigg J, McIver M, Jüppner H, Alexander SI. 2005 Multicystic dysplastic kidney and variable phenotype in a family with a novel deletion mutation of *PAX2*. *J. Am. Soc. Nephrol.* **16**, 2754–2761. (doi:10.1681/ASN.2005030239)
- Bower M *et al.* 2012 Update of *PAX2* mutations in renal coloboma syndrome and establishment of a locus-specific database. *Hum. Mutat.* **33**, 457–466. (doi:10.1002/humu.22020)
- Hero I. 1990 Optic fissure closure in the normal cinnamon mouse. *Invest. Ophthalmol. Vis. Sci.* **31**, 197–216.
- Masai I *et al.* 2003 N-cadherin mediates retinal lamination, maintenance of forebrain compartments and patterning of retinal neurites. *Development* **130**, 2479–2494. (doi:10.1242/dev.00465)
- Chen S, Lewis B, Moran A, Xie T. 2012 Cadherin-mediated cell adhesion is critical for the closing of the mouse optic fissure. *PLoS ONE* **7**, e51705. (doi:10.1371/journal.pone.0051705)
- James A, Lee C, Williams AM, Angileri K, Lathrop KL, Gross JM. 2016 The hyaloid vasculature facilitates basement membrane breakdown during choroid fissure closure in the zebrafish eye. *Dev. Biol.* **419**, 262–272. (doi:10.1016/j.ydbio.2016.09.008)
- Wu MY, Hill CS. 2009 TGF- β Superfamily signaling in embryonic development and homeostasis. *Dev. Cell* **16**, 329–343. (doi:10.1016/j.devcel.2009.02.012)
- Zhang J, Tian X-J, Xing J. 2016 Signal transduction pathways of EMT Induced by TGF- β , SHH, and WNT and their crosstalks. *J. Clin. Med.* **5**, 41. (doi:10.3390/jcm5040041)
- Thiery JP, Acloque H, Huang RYJ, Nieto MA. 2009 Epithelial-mesenchymal transitions in development and disease. *Cell* **139**, 871–890. (doi:10.1016/j.cell.2009.11.007)
- Mercado-Pimentel ME, Runyan RB. 2007 Multiple transforming growth factor-beta isoforms and receptors function during epithelial-mesenchymal cell transformation in the embryonic heart. *Cells Tissues Organs* **185**, 146–156. (doi:10.1159/000101315)
- Johansson J *et al.* 2013 MiR-155-mediated loss of *C/EBP β* shifts the TGF- β response from growth inhibition to epithelial-mesenchymal transition, invasion and metastasis in breast cancer. *Oncogene* **32**, 5614–5624. (doi:10.1038/onc.2013.322)
- Verrecchia F, Mauviel A. 2007. Transforming growth factor- β and fibrosis. *World J. Gastroenterol.* **13**, 3056–3062. (doi:10.3748/wjg.v13.i22.3056)
- Izumi N *et al.* 2006 BMP-7 opposes TGF- β 1-mediated collagen induction in mouse pulmonary myofibroblasts through *Id2*. *Am. J. Physiol. Lung Cell Mol. Physiol.* **290**, L120–L126. (doi:10.1152/ajplung.00171.2005)
- Zeisberg M, Hanai J, Sugimoto H, Mammoto T, Charytan D, Strutz F, Kalluri R. 2003 BMP-7 counteracts TGF- β 1-induced epithelial-to-mesenchymal transition and reverses chronic renal injury. *Nat. Med.* **9**, 964–968. (doi:10.1038/nm888)

31. Wang S, Hirschberg R. 2003 BMP7 antagonizes TGF- β -dependent fibrogenesis in mesangial cells. *Am. J. Physiol. Renal Physiol.* **284**, F1006–F1013. (doi:10.1152/ajprenal.00382.2002)
32. Wang S, Hirschberg R. 2004 Bone morphogenetic protein-7 signals opposing transforming growth factor β in mesangial cells. *J. Biol. Chem.* **279**, 23 200–23 206. (doi:10.1074/jbc.M311998200)
33. Sanford LP, Ormsby I, Gittenberger-de Groot AC, Sariola H, Friedman R, Boivin GP, Cardell EL, Doetschman T. 1997 TGF β 2 knockout mice have multiple developmental defects that are non-overlapping with other TGF β knockout phenotypes. *Development* **124**, 2659–2670.
34. Rahhal B, Heermann S, Ferdinand A, Rosenbusch J, Rickmann M, Kriegelstein K. 2009 *In vivo* requirement of TGF- β /GDNF cooperativity in mouse development: focus on the neurotrophic hypothesis. *Int. J. Dev. Neurosci.* **27**, 97–102. (doi:10.1016/j.ijdevneu.2008.08.003)
35. Pichel JG *et al.* 1996 Defects in enteric innervation and kidney development in mice lacking GDNF. *Nature* **382**, 73–76. (doi:10.1038/382073a0)
36. Moore MW *et al.* 1996 Renal and neuronal abnormalities in mice lacking GDNF. *Nature* **382**, 76–79. (doi:10.1038/382076a0)
37. Sánchez MP, Silos-Santiago I, Frisén J, He B, Lira SA, Barbacid M. 1996 Renal agenesis and the absence of enteric neurons in mice lacking GDNF. *Nature* **382**, 70–73. (doi:10.1038/382070a0)
38. Hellmich HL, Kos L, Cho ES, Mahon KA, Zimmer A. 1996 Embryonic expression of glial cell-line derived neurotrophic factor (GDNF) suggests multiple developmental roles in neural differentiation and epithelial-mesenchymal interactions. *Mech. Dev.* **54**, 95–105. (doi:10.1016/0925-4773(95)00464-5)
39. Fuchshofer R, Birke M, Welge-Lüssen U, Kook D, Lutjens-Drecoll E. 2005 Transforming growth factor- β 2 modulated extracellular matrix component expression in cultured human optic nerve head astrocytes. *Invest. Ophthalmol. Vis. Sci.* **46**, 568–578. (doi:10.1167/iovs.04-0649)
40. Heldin C-H, Miyazono K, ten Dijke P. 1997 TGF- β signalling from cell membrane to nucleus through SMAD proteins. *Nature* **390**, 465–471. (doi:10.1038/37284)
41. Dennler S, Itoh S, Vivien D, ten Dijke P, Huet S, Gauthier JM. 1998 Direct binding of Smad3 and Smad4 to critical TGF β -inducible elements in the promoter of human plasminogen activator inhibitor-type 1 gene. *EMBO J.* **17**, 3091–3100. (doi:10.1093/emboj/17.11.3091)
42. Lin AH, Luo J, Mondschein LH, ten Dijke P, Vivien D, Contag CH, Wyss-Coray T. 2005 Global analysis of Smad2/3-dependent TGF- β signaling in living mice reveals prominent tissue-specific responses to injury. *J. Immunol.* **175**, 547–554. (doi:10.4049/jimmunol.175.1.547)
43. Inman GJ, Nicolás FJ, Callahan JF, Harling JD, Gaster LM, Reith AD, Laping NJ, Hill CS. 2002 SB-431542 is a potent and specific inhibitor of transforming growth factor- β superfamily type I activin receptor-like kinase (ALK) receptors ALK4, ALK5, and ALK7. *Mol. Pharmacol.* **62**, 65–74. (doi:10.1124/mol.62.1.65)
44. Laping NJ *et al.* 2002 Inhibition of transforming growth factor (TGF)- β 1-induced extracellular matrix with a novel inhibitor of the TGF- β type I receptor kinase activity: SB-431542. *Mol. Pharmacol.* **62**, 58–64. (doi:10.1124/mol.62.1.58)
45. Schwend T, Loucks EJ, Ahlgren SC. 2010 Visualization of Gli activity in craniofacial tissues of hedgehog-pathway reporter transgenic zebrafish. *PLoS ONE* **5**, e14396. (doi:10.1371/journal.pone.0014396)
46. Jinnin M, Ihn H, Tamaki K. 2006 Characterization of SIS3, a novel specific inhibitor of Smad3, and its effect on transforming growth factor- β 1-induced extracellular matrix expression. *Mol. Pharmacol.* **69**, 597–607. (doi:10.1124/mol.105.017483)
47. Koshiba-Takeuchi K *et al.* 2000 Tbx5 and the retinotectum projection. *Science* **287**, 134–137. (doi:10.1126/science.287.5450.134)
48. Sasagawa S, Takabatake T, Takabatake Y, Muramatsu T, Takeshima K. 2002 Axes establishment during eye morphogenesis in *Xenopus* by coordinate and antagonistic actions of BMP4, Shh, and RA. *Genesis* **33**, 86–96. (doi:10.1002/gene.10095)
49. Mui SH, Kim JW, Lemke G, Bertuzzi S. 2005 Vax genes ventralize the embryonic eye. *Genes Dev.* **19**, 1249–1259. (doi:10.1101/gad.1276605)
50. Behesti H, Holt JKL, Sowden JC. 2006 The level of BMP4 signaling is critical for the regulation of distinct T-box gene expression domains and growth along the dorso-ventral axis of the optic cup. *BMC Dev. Biol.* **6**, 62. (doi:10.1186/1471-213X-6-62)
51. Müller II, Knapik EW, Hatzopoulos AK. 2006 Expression of the protein related to Dan and Cerberus gene—*prdc*—during eye, pharyngeal arch, somite, and swim bladder development in zebrafish. *Dev. Dyn.* **235**:2881–2888. (doi:10.1002/dvdy.20925)
52. See AW-M, Clagett-Dame M. 2009 The temporal requirement for vitamin A in the developing eye: mechanism of action in optic fissure closure and new roles for the vitamin in regulating cell proliferation and adhesion in the embryonic retina. *Dev. Biol.* **325**, 94–105. (doi:10.1016/j.ydbio.2008.09.030)
53. Barbieri AM *et al.* 2002 Vax2 inactivation in mouse determines alteration of the eye dorsal-ventral axis, misrouting of the optic fibres and eye coloboma. *Development* **129**, 805–813.
54. Proetzl G, Pawlowski SA, Wiles MV, Yin M, Boivin GP, Howles PN, Ding J, Ferguson MW, Doetschman T. 1995 Transforming growth factor-beta 3 is required for secondary palate fusion. *Nat. Genet.* **11**, 409–414. (doi:10.1038/ng1295-409)
55. Taya Y, O'Kane S, Ferguson MW. 1999 Pathogenesis of cleft palate in TGF-beta3 knockout mice. *Development* **126**, 3869–3879.
56. Zajzon K, Pröls F, Heermann S. 2013 Concerted interaction of TGF- β and GDNF mediates neuronal differentiation. *Neuroreport* **24**, 704–711. (doi:10.1097/WNR.0b013e328363f75c)
57. Peterziel H, Paech T, Strelau J, Unsicker K, Kriegelstein K. 2007 Specificity in the crosstalk of TGFbeta/GDNF family members is determined by distinct GFR alpha receptors. *J. Neurochem.* **103**, 2491–2504. (doi:10.1111/j.1471-4159.2007.04962.x)
58. Kriegelstein K, Henheik P, Farkas L, Jaszai J, Galter D, Krohn K, Unsicker K. 1998 Glial cell line-derived neurotrophic factor requires transforming growth factor-beta for exerting its full neurotrophic potential on peripheral and CNS neurons. *J. Neurosci.* **18**, 9822–9834.
59. Insua MF, Garelli A, Rotstein NP, German OL, Arias A, Politi LE. 2003 Cell cycle regulation in retinal progenitors by glia-derived neurotrophic factor and docosahexaenoic acid. *Invest. Ophthalmol. Vis. Sci.* **44**, 2235–2244. (doi:10.1167/iovs.02-0952)
60. Morcillo J, Martínez-Morales JR, Trousse F, Fermin Y, Sowden JC, Bovolenta P. 2006 Proper patterning of the optic fissure requires the sequential activity of BMP7 and SHH. *Development* **133**, 3179–3190. (doi:10.1242/dev.02493)
61. Zode GS, Clark AF, Wordinger RJ. 2009 Bone morphogenetic protein 4 inhibits TGF- β 2 stimulation of extracellular matrix proteins in optic nerve head cells: role of gremlin in ECM modulation. *Glia* **57**, 755–766. (doi:10.1002/glia.20803)
62. Al Chawa T *et al.* 2014 Nonsyndromic cleft lip with or without cleft palate: increased burden of rare variants within Gremlin-1, a component of the bone morphogenetic protein 4 pathway. *Birth Defects Res. Part A Clin. Mol. Teratol.* **100**, 493–498. (doi:10.1002/bdra.23244)
63. Thisse *et al.* 2001 Expression of the zebrafish genome during embryogenesis. ZFIN publication. See <https://zfin.org/ZDB-PUB-010810-1> (accessed 19 May 2017).
64. Thisse B, Thisse C. 2004 Fast release clones: a high throughput expression analysis. ZFIN publication. See <https://zfin.org/ZDB-PUB-040907-1> (accessed 19 May 2017).
65. Thisse C, Thisse B. 2005 High throughput expression analysis of ZF-Models Consortium clones. ZFIN publication. See <https://zfin.org/ZDB-PUB-051025-1> (accessed 19 May 2017).
66. Shawi M, Serluca FC. 2008 Identification of a BMP7 homolog in zebrafish expressed in developing organ systems. *Gene Expr. Patterns* **8**, 369–375. (doi:10.1016/j.gexp.2008.05.004)
67. Khokha MK, Yeh J, Grammer TC, Harland RM. 2005 Depletion of three BMP antagonists from Spemann's organizer leads to a catastrophic loss of dorsal structures. *Dev. Cell* **8**, 401–411. (doi:10.1016/j.devcel.2005.01.013)
68. Eggen BJ, Hemmati-Brivanlou A. 2001 BMP antagonists and neural induction. In *eLS*. Chichester, UK: John Wiley & Sons, Ltd. <http://www.els.net> (doi:10.1038/npg.els.0000805).
69. Stottmann RW, Anderson RM, Klingensmith J. 2001 The BMP antagonists Chordin and Noggin have essential but redundant roles in mouse mandibular outgrowth. *Dev. Biol.* **240**, 457–473. (doi:10.1006/dbio.2001.0479)

70. Pogoda H-M, Meyer D. 2002 Zebrafish Smad7 is regulated by Smad3 and BMP signals. *Dev. Dyn.* **224**, 334–349. (doi:10.1002/dvdy.10113)
71. Zhang R, Huang H, Cao P, Wang Z, Chen Y, Pan Y. 2013 Sma- and Mad-related protein 7 (Smad7) is required for embryonic eye development in the mouse. *J. Biol. Chem.* **288**, 10 275–10 285. (doi:10.1074/jbc.M112.416719)
72. Labbé E, Silvestri C, Hoodless PA, Wrana JL, Attisano L. 1998 Smad2 and Smad3 positively and negatively regulate TGF β -dependent transcription through the Forkhead DNA-binding protein FAST2. *Mol. Cell* **2**, 109–120. (doi:10.1016/S1097-2765(00)80119-7)
73. Chen X, Rubock MJ, Whitman M. 1996 A transcriptional partner for MAD proteins in TGF- β signalling. *Nature* **383**, 691–696. (doi:10.1038/383691a0)
74. Jonk LJ, Itoh S, Heldin CH, ten Dijke P, Kruijer W. 1998 Identification and functional characterization of a Smad binding element (SBE) in the JunB promoter that acts as a transforming growth factor- β , activin, and bone morphogenetic protein-inducible enhancer. *J. Biol. Chem.* **273**, 21 145–21 152. (doi:10.1074/jbc.273.33.21145)
75. R Core Team. 2014 *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. See <http://www.R-project.org/>
76. Reimand J, Arak T, Adler P, Kolberg L, Reisberg S, Peterson H, Vilo J. 2016 g:Profiler-a web server for functional interpretation of gene lists (2016 update). *Nucleic Acids Res.* **44**, W83–W89. (doi:10.1093/nar/gkw199)
77. Chomczynski P, Sacchi N. 2006 The single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction: twenty-something years on. *Nat. Protoc.* **1**, 581–585. (doi:10.1038/nprot.2006.83)
78. Schmittgen TD, Livak KJ. 2008 Analyzing real-time PCR data by the comparative Ct method. *Nat. Protoc.* **3**, 1101–1108. (doi:10.1038/nprot.2008.73)
79. Kwan KM *et al.* 2007 The Tol2kit: a multisite gateway-based construction kit for Tol2 transposon transgenesis constructs. *Dev. Dyn.* **236**, 3088–3099. (doi:10.1002/dvdy.21343)
80. Kirchmaier S, Höckendorf B, Möller EK, Bornhorst D, Spitz F, Wittbrodt J. 2013 Efficient site-specific transgenesis and enhancer activity tests in medaka using PhiC31 integrase. *Development* **140**, 4287–4295. (doi:10.1242/dev.096081)
81. Dogra D, Ahuja S, Kim HT, Rasouli SJ, Stainier DYS, Reischauer S. 2017 Opposite effects of Activin type 2 receptor ligands on cardiomyocyte proliferation during development and repair. *Nat. Com.* **8**, 1902. (doi: 10.1038/s41467-017-01950-1)
82. Casari A, Schiavone M, Facchinello N, Vettori A, Meyer D, Tiso N, Moro E, Argenton F. 2014 A Smad3 transgenic reporter reveals TGF-beta control of zebrafish spinal cord development. *Dev. Biol.* **396**, 81–93. (doi:10.1016/j.ydbio.2014.09.025)
83. Quiring R, Wittbrodt B, Henrich T, Ramialison M, Burgdorf C, Lehrach H, Wittbrodt J. 2004 Large-scale expression screening by automated whole-mount in situ hybridization. *Mech. Dev.* **121**, 971–976. (doi:10.1016/j.mod.2004.03.031)