A Portable IoT NIR Spectroscopic System to Analyze

the Quality of Dairy Farm Forage

Guillermo Rego, ¹ Francisco Ferrero, ¹ Marta Valledor, ¹ Juan Carlos Campo, ¹ Sergio Forcada, ² Luis J. Royo, ² Ana Soldado ²

¹ Dept. of Electrical, Electronic, Computers, and System Engineering University of Oviedo, Campus of Gijón, Spain

² Dept. of Nutrition, Grasslands and Forage, Regional Institute for Research and Agro-Food Development, Villaviciosa, Spain

Abstract

Having rapid and reliable knowledge of the quality of dairy farm forage is crucial for technicians and producers. However, the necessary equipment to fulfil these requirements is oftentimes expensive and complex to use. This work presents a portable instrument system to analyze the nutritional values of dairy farm forage using near-infrared spectrometry (NIRS) techniques. Using the Internet of Things (IoT) tools, data are sent to the cloud for processing; following this, they are accessible to any device. To analyze the nutritional values of dairy farm forage a chemometric model and implemented the instrument to understand the relationship between the measured spectrum and the concentration of the substances of interest was developed. The quality of the proposed methodology was validated by comparing reference and NIR data by using the statistic Standard Error of Calibration.

Keywords

Food quality, NIRS, spectroscopy, chemometrics, Internet of Things (IoT), cloud computing.

1. Introduction

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

Possessing quick and reliable knowledge of the nutritive values of dairy farm forage is crucial for technicians and producers. Most instrumental techniques employed for quality evaluation are time-intensive, expensive, and involve a considerable amount of manual work. There is growing interest in developing simpler and faster instrumental devices for determining the physical and chemical characteristics in agricultural system samples. In spectroscopic analysis, an alternative for raw sample analysis is near-infrared spectroscopy (NIRS), which has taken its place among other proven spectroscopic tools for determining the chemical and physical properties of foods. The NIRS absorption originated from the overtones of CH, NH, OH, and SH stretching vibrations and from stretching-bending combinations involving these groups. The near-infrared spectrum is located between the infrared and visible spectrum, from 2500 nm to 800 nm (4000–12500 cm⁻¹) (Ozaki et al., 2007; Williams et al., 2001), and the spectral signal can be used for advanced analytics. However, this technology traditionally has been reserved as an expensive laboratory instrument for experts. Because of the numerous publications that support the efficacy of analytical applications that use NIRS for the quantification of quality and safety parameters in agri-food (Decruyenaere et al., 2009; Baeten et al., 2008; De la Haba et al., 2007; Fernández Pierna et al., 2006; Okparanma et al., 2013; Volkers et al., 2003; Hermida et al., 2005), researchers are working on developing new portable NIRS instruments.

Related to feed analysis, numerous authors have reported using NIRS to conduct rapid analysis of feed nutritional quality (Nie et al. 2008; Huang et al. 2008; van Barneveld et al. 2018). NIRS holds real potential for routine on-site analyses, for example at the point of feed delivery or for use in stock feed manufacturing (Yan et al., 2018).

The NIRS researcher, user, and engineer must give special attention to sample presentation devices that directly govern the quality of spectra themselves. There is a technique available for any type of liquid, slurry, powdered or solid sample. Radiation interacting with a sample may be absorbed, transmitted, or reflected. For raw forage analysis, the solid sample is used as a diffuse reflectance because most of the incident radiation is reflected. In this mode, the incident radiation

illuminates perpendicularly to the sample surface. Normally, the incident radiation cannot reach a deeper position in a sample because of high absorption or multiple scattering (Ozaki et al., 2007).

NIR spectra contain a lot of physical and chemical molecule information. However, this information cannot always be extracted straightforwardly from the spectra because the NIR spectra consists of a number of bands arising from overtones and combination modes that overlap heavily with one another. Additionally, multicollinearity is quite strong in this region; therefore, powerful chemometric techniques are essential for allowing reliable extraction of relevant information hidden in the NIR spectral data. Chemometrics covers all methods of multivariable calibration, including spectral data preprocessing, and calibration model development for qualitative and quantitative analyses.

Advances in NIR spectroscopy have made this technology one of the most efficient tools for determining feed nutritive parameters. Based on the type of detector, the portable NIRS can be an array detector or single-detector instrument. Comparing both detectors, the price for a single one is lower and thus reduces the hardware costs. These reasons have facilitated the development of instruments with single detectors (Olakunle et al., 2018). Besides the price and portability of the miniaturized NIRS systems to carry out on-site and in-field measurements, there is another problem to solve, one related to data transference and management. Existing NIRS instruments cannot extract the spectra data easily because they do not usually have the appropriate connectivity. This causes a problem when it comes to following the evolution of the data. This information can be useful for technicians, producers, or administration responsible for food quality and safety. Having data history is always an added benefit that can provide valuable information about what is happening on the farm or factory. In this sense, advances in communication technologies and smartphones open a range of new possibilities that could be applied to develop a solution fitting the desired conditions. Thus, Olakunle et al. discussed in detail the benefits and challenges of Internet of Things (IoT) technologies and data analytics in agriculture. Liu et al. (Liu et al., 2019) developed and implemented a modern agriculture IoT system for the cloud, whereas Crocombe (Crocombe, 2018) provided detailed information about the technologies used in portable/miniaturized spectroscopy, discussing their applications and emerging fields. A review of recent smartphone spectroscopy systems is provided in (Rateni et al., 2017).

In summary, the main contributions of this work are the following:

- From a NIR evaluation module, a high-performance portable NIR spectrometer was developed.
- A chemometric model into the instrument to extract relevant information about forage quality was implemented.
 - IoT capability were included in the developed NIR spectrometer to process and communicate data everywhere and all the time, with different devices with access to the Internet.

The remainder of this paper is organized as follows: Section II presents the materials used in this work. Section III describes the implementation of the IoT NIR measurement system. Section IV is devoted to the chemometrics analysis. Cloud computing is addressed in section V. Finally, Section VI contains the conclusion.

2. Materials

Generally, an NIRS system comprises four components: light source, light-isolating mechanisms, detector, and sampling device (Wang et al. 2007). Most of the instruments utilize tungsten filament halogen lamps as the light source. This is because they are an effective compromise between performance and cost. This type of lamp is particularly suitable for use in low voltage settings. The light source must be close to the sample to light it up with strong intensity. The employed technology for the wavelengths selection greatly influences the performance of the instrument.

Regarding portable applications in food sector instruments equipped with a diffraction grating and an array detector, the Si-diode array or InGaAs-diode array have seen increased use due to their compact size, versatility, robustness, and low cost. Fig. 1 shows the spectrometer approach proposed by Texas Instruments (TI) (Texas Instruments, 2016). The light source is first collimated and then sent to the diffraction grating to split into different wavelengths. The focusing mirror collects the diffracted radiation and sends it to a digital mirror array (DMD) instead of an array detector. The DMD consists of an array of hundreds of thousands of micromirrors.

However, when activating or not activating the mirror columns, only the desired wavelengths are reflected by the detector. This programmable architecture allows the use of a high-performance, cost-effective single element detector—for example a photonic device based on InGaAs. This approach has a small form factor suited for field analysis, which provides mechanical stability. In this work, the TI NIRscan Nano evaluation module is used (Texas Instruments, 2017) (see Fig. 2a). It is a compact battery-operated evaluation module for portable NIRS solutions. It supports Bluetooth Low Energy (BLE) to enable mobile measurements for hand-held spectrometers. This module operates by illuminating the test sample at an angle so that specular reflections are not collected. At the same time, it gathers and focuses diffuse reflections through the slit. The embedded processor commands the DMD controller turn on only the precise mirrors, which are illuminated by the specific light wavelengths to be measured at each instant of time. The easy programmability of the processor allows users to command specific column widths or other patterns for their spectroscopy requirements to achieve high signal-to-noise ratio values. This architecture enables adaptive scanning techniques, which are not possible with array detectors for example, chemometric methods with multiple patterns to look for physical and chemical substances. A microcontroller board (LOLIN D32) is attached over the spectrometer module (see Fig. 2b) to control the servomotor (MG90S) that moves the sample holder. A DC-DC converter (Pololu U1V10F5) is used to convert the 3.7 V from the battery to 5 V to power the servomotor.

131

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

3. IoT Measurement System

Fig. 3 shows the general scheme of the proposed measurement system. It consists of the NIRscan Nano module and a microcontroller board based on the well-known ESP32 module, which is responsible for turning the servomotor (M) where the sample holder is placed. The spectrometer will make several measurements when the sample holder is rotating, which ensures the measurements are as homogeneous as possible. These data can be sent to a mobile application via Bluetooth, which is integrated into the NIRscan. There is also the option of sending them to the computer via a USB connection.

Due to the limitation of storage and computing capability of the local system on chip, some researchers use a limited chemometrics model for portable spectra instruments. However, in the proposed instrument, spectra data are sent to the cloud, where, with the rapid progress of IoT and cloud computing, along with the development of machine-learning algorithms, real-time applications can be guaranteed. Once the model or models are deployed on the cloud, there is no limitation on storage or computing capability. In addition, the results are accessible from other mobile devices with Internet access.

The small size of the sample window (10 mm x 10 mm) makes it difficult to obtain reproducible spectra analyzing raw samples with a particle size larger than the instrument sample window when measuring them directly. To avoid this drawback, a semi-circular sample holder was attached to the spectrometer, as shown in Fig. 4. This holder rotates and, in each position, 10 measurements are made. The microcontroller board has two buttons, one to start the measurements and another to set the load position of the sample holder.

When the start button is pressed, the servomotor begins to turn, stopping at different positions where the measurements are made. In this research work, it was optimized the number of positions to be scanned per sample going from 5 to 20 positions. The best results were obtained when scanning 10 points per sample. Once the sample holder reaches the 180° position, it returns to the initial position, where it waits to make a new measurement. When the load button is pressed, the

servomotor goes to an intermediate position of about 90°, facilitating the loading and unloading of the sample. When the device is not performing either of these two actions, the microcontroller will go into sleep mode to minimize power consumption. The movement of the alfalfa inside the holder does not affect the measure because the quantity of alfalfa that is introduced, is large enough so that the movement of it inside the sample holder is minimal. What is sought when performing the 10 measurements is to obtain the most homogeneous results possible in the sample, so that even if it moved, it would not affect the result since the exact measurement points are not really important, but the important thing is to cover the largest sample area.

Autonomy is fundamental in any portable IoT device. In this work a 3.7 V, 1800 mAh Lithium polymer battery is used to power the system, providing an estimated autonomy of 15 hours when it is at rest. A 10 k Ω NTC thermistor is required to safely charge the battery and monitor its temperature. When the measurement is being made, the spectrometer alone consumes 500 mA, but the measurement time is only about 16 s. Therefore, if the user makes an average of 10 measurements per hour, the autonomy of the prototype will be 12 hours (if it is on all the time). Texas Instruments provides free software and firmware downloads to give developers flexibility and advanced control of the spectrometer.

4. Chemometric Analysis

Chemometrics are a wide range of statistical and mathematical methods for extracting useful information from the NIR spectra. These methods use a multivariable calibration in which the wide spectrum is considered to establish a relationship between spectra data and analytical parameter quantification. Fig. 5 shows the process to develop the chemometric model for the quantification of nutritive forage value.

A. Sample Selection

The first task of this process is the selection of the sample group for calibration, which must be well defined statistically, including a wide variability of the type of samples. In this work, 90 samples were collected from different farms in Asturias Province (north Spain) during 2018–2019

and analyzed using the NIRS designed instrument and reference procedures. For reference data, we performed the chemical analysis using traditional analytical methodologies: Crude Protein (CP) through Kjeldahl analysis, Neutral Detergent Fiber (NDF), and Acid Detergent Fiber (ADF) through Van Soest analysis (Van Soest et al., 1991). Table 1 provides the range, mean, and standard deviation of the reference analysis. As can be seen, it was observed a wide variability when determining fiber parameters (Standard deviation >5) due to different maturity stages of the plant; however, it was noted a lower standard deviation for the crude protein parameter. The protein content of alfalfa as cut varies according to the cut and vegetative stage of the crop and the leaf content after the drying procedure.

Table 1. Statistic for Nutritive Value of Alfalfa Samples (N = 90)

Analytical Methodologies	Range (%)	Mean	Standard Deviation
СР	7.19 - 17.27	14.49	1.61
ADF	20.15 - 39.24	29.22	4.00
NDF	29.24 - 60.44	39.36	5.40

CP: Crude Protein; ADF: Acid Detergent Fibre; NDF: Neutral Detergent Fibre.

B. Spectral Acquisition

To scan a sample, a scan configuration must be created. Two scan configurations come preloaded in factory settings: "Column" and "Hadamard." Column selects one wavelength at a time. The Hadamard scan creates a set with several wavelengths multiplexed at a time and then decodes the individual wavelengths. Fig. 6 displays the spectrum plot and the scan configuration parameters. A scan can be divided into 1–5 sections, being configured in each section: the method (Column or Hadamard), spectral range (start and end wavelength), digital resolution (wavelength points captured into the defined spectral range), exposure time (between 0.635 – 60.960 ms) and the number of scans per sample (in this work, 10 scans in 10 different points of the sample).

The NIR spectrum contains information about the major X-H chemical bonds in an agricultural product such as lucerne. The spectrum is dependent on all the functional groups that

absorb NIR radiation, and this information has important implications in agricultural products due to the relationship between NIR vibration bands of functional groups and nutritional parameters. Moreover, the reference bands related to crude protein are associated to N-H vibrations in the spectra regions around 1470, 1500 to 1530 and 1640 to 1680 nm.

For the major fiber components, the tentative bands assignments are associated to structures of cellulose, pectins and lignin, with absorbance at 1170, 1420 and 1490nm. Lucerne spectra collected with the developed device are shown in Fig. 7 (Burns and Ciurczak, 2008).

C. Spectral Preprocessing

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

An important effect related to analysing intact samples by NIR is the particle size, being desirable to try to eliminate or reduce it, using a variety of mathematical approaches for spectra preprocessing. The aim of this step is to improve the signal-to-noise ratio, removing the redundant information contained in the spectra. There are several methods for preprocessing spectra data, such as averaging all the individual spectra collected for one sample or using smoothing or derivative procedures. The first one consists of performing a median of all the measures. The result is a single spectrum of each sample with a reduced thermal noise. Another effect to be minimized is scattering: in NIRS, the reflected energy is a mixture of diffuse and specular reflections, which are dependent on the scattering nature and absorption characteristics of the sample (Bertrand et al. 2000). This makes the baseline/intensity of each sample vary, introducing challenges when making a model. To solve this issue, transformations such as the multiplicative scatter correction (Barnes et al. 1989) and the standard normal variate correction (SNV) (aes et al., 2004) are common in NIRS chemometrics. In this work, the SNV was applied to each spectrum individually and calculated the mean and standard deviation of each point. To calculate the SNV spectra, the average at that point (μ) is subtracted from each point and divided by the standard deviation (σ) . The correction can be seen in Figs. 7 and 8.

To improve the quality of the spectra, a Savitzky–Golay filter was used. This is a low-pass filter which applies least squares smoothing to reduce noise while maintaining the shape and height of waveform peaks, which is why it is widely used in spectrometry. In this work, a window

size of 19, a polynomial of order 3, and a derivative of order 2 were used. The result of the pretreatment can be seen in Fig. 9. In addition, to improve model results, the wavelengths that do not provide information from the model were analyzed and removed.

When applying SNV and Detrend on spectra population (Fig. 7), the effects of particle size are compensated and shift of the base line is removed or minimized (see Fig. 8).

D. Calibration Model

The next step after preprocessing is the calibration of the model. It consists of obtaining a correlation between the reference method and the spectral data. The most widely used regression methods in NIRS are multiple linear regression (Martens et al. 1989), principal components regression, and partial least squares regression (PLSR) (Wold et al., 2001). This last method can improve the precision of the model parameters with the increase in number of relevant variables and observations, and it is widely used in NIRS calibration developments. To attempt this work, a PLSR model has been employed using all the data included in the spectra. To select the best equations, the following statistics have been considered: standard error of calibration (SEC) and determination coefficient of calibration (R^2_C). The first one (R^2_C) establishes a correlation between the analytical data obtained in the laboratory and those predicted by the calibration (R^2_C). Mathematically,

255
$$R^{2} = 1 - \frac{\sum_{i=1}^{n} (y_{i} - \hat{y}_{i})^{2}}{\sum_{i=1}^{n} (y_{i} - \bar{y})^{2}}$$
 (1)

where n is the number of samples used in the calibration, y_i refers to the reference values, \hat{y}_i is the value predicted by the model, and \hat{y}_i is the average reference value.

The second parameter is calculated using the formula:

259
$$SEC = \sqrt{\frac{1}{n} \cdot \sum_{i=1}^{M} (y_i - \hat{y}_i)^2}$$
 (2)

where \hat{y}_i and y_i are the predicted and measured values for sample ith. This value provides the averages of expected typical uncertainty for the prediction of future samples.

Table 2 shows the best statistics of chemometric models for predicting nutritive values in alfalfa samples. These characteristics were selected according to the lowest standard error of calibration and the lowest error of prediction. As can be seen, when increasing variability in calibration, population R^2_C is increased. For CP with a lower variability (SD = 1.61), the R^2_C is 0.516. To evaluate the statistical data of developed models, this work focused on the SEC and the ability of being useful to farms for feeding management.

The standard errors in the developed models were obtained according to Commission Regulation (EU) 2017/2279 of 11 December 2017 amending Annexes II, IV, VI, VII, and VIII to Regulation (EC) No 767/2009 of the European Parliament and to the ISO 13906 and 16472 related to these nutritive parameters.

Table 2. Statistics of Chemometric Models to Predict Nutritive Parameters of Alfalfa Samples

Analytical Methodologies	$ m R^2_{C}$	SEC
СР	0.516	2.11
ADF	0.742	3.96
NDF	0.704	8.03

CP: Crude Protein; ADF: Acid Detergent Fibre; NDF: Neutral Detergent Fibre; R²_C: Determination coefficient for calibration; SEC: Standard Error of Calibration; RPD: Standard deviation/SEC.

5. Cloud Computing

According to the schema in Fig. 5, after performing the measurement, the spectrometer sends the data in the "csv" format to the computer via BLE. Once there, the spectra data is sent to the cloud, where the statistical treatment presented in Section IV was carried out. Using the Python language, a dashboard to display the data and allow users to upload and download the spectra was developed.

The provider used to host the application is Amazon Web Services. Fig. 10 shows the dashboard structure on the cloud. To facilitate its deployment in other platforms, the developed application runs inside a Docker container. The user, using a web browser, where the dashboard is displayed, communicates with the cloud through the free and open source software, Nginx. This works as a web server and allows users to communicate with the application itself, giving the system a layer so that the data are not directly exposed. After that, a Python WSGI HTTP Server for UNIX (Gunicorn) is find. The developed application utilizes this software. The application was developed using Dash, which is a framework for building analytical web applications. The data spectra are stored in a database, a nonrelational and open source database called MongoDB.

Finally, the pre-processed spectra data (see C. Spectral Preprocessing section) are displayed in a dashboard where the user can see the results and the graphics generates, as can be seen in Fig. 11. Deploying the entire system in the cloud allows us to adapt new volumes of data in the future with relative ease. It is possible also do so in such a way that any authorized person can access it from anywhere in the world if they have access to the Internet and not depend on the construction and maintenance of their own servers, which makes a very low initial cost.

6. Conclusion

This work utilizes an affordable IoT portable NIRS system to analyze the nutritional parameters of dairy farm forage. This system can be useful for the owners or technicians responsible for the nutritional state of animals in farms. The proposed instrumentation allows users to evaluate the quality of the forage, increase sampling without incurring a cost, obtain results in real time, and make it possible to make quick decisions, avoiding delays related to carrying samples from the farm to the laboratory. Moreover, specialized training is not required for users of this instrument. Thanks to the storage and processing in the cloud, the data are accessible from any site with Internet access, facilitating the visualization and use of the data for making decisions. The potential offered by cloud computing suggests this system would be useful not only for agriculture monitoring but also for environmental and biomedical sensing.

312	Acknowledgments
313	This work was supported by the Spanish government (Ministry of Economy, Industry, and
314	Competitiveness) through the RTA2015-00061-C02-02 project and from the European Union and
315	Principality of Asturias through the GRUPIN-IDI/2018/000166 and GRUPIN IDI/2018/000237
316	(PCTI and FEDER) projects.
317	
318	References
319	Ozaki, Y., Fred McClure, W., Christy, A.A., 2006. Near-Infrared Spectroscopy in food science
320	and technology. John Wiley & Sons, Inc.
321	Williams, P., Norris, K. (Eds). 2001. Near infrared technology in the agricultural and food
322	industry, second edition.
323	Decruyenaere, V., Lecomte, Ph., Demarquilly, C., Aufrere, J., Dardenne, P., Stilmant, D.
324	Buldgen, A. 2009. Evaluation of green forage intake and digestibility in ruminants using near
325	infrared reflectance spectrscopy (NIRS): Developing a global calibration. Animal Feed
326	Science Technololy, 148, 138-156.
327	Baeten, V., Manley, M., Fernandez Pierna, J.A., Downey, G., Dardenne, P. 2008. Spectrometric
328	Technique: Fourier Transform Mear-infrared (FT-NIR) Spectroscopy. In: Modern
329	Techniques for Food Authentication, Sun, Da-Wen Sun. Dublin - UK, Elsevier. 117-147.
330	De la Haba, M.J., Fernández Pierna, J.A., Fumière, O., Garrido-Varo, A., Guerrero J.E., Pérez-
331	Marín D.C., Dardenne P., Baeten V. 2007. Discrimination of fish bones from other animal
332	bones in the sedimented fraction of compound feeds by near infrared microscopy. J. of NIRS
333	15, 81-88.

Fernández Pierna, J.A., Baeten, V., Dardenne, P. Screening of compound feeds using NIR

hyperspectral data. 2006. Chemometrics and Intelligent Laboratory Systems, 84, 114-118.

- Fernández-Ibáñez, V., Soldado A., Vicente, F., Martínez-Fernández, A., De La Roza-Delgado, B.
- 337 2008. Particle size optimisation in development of near infrared microscopy methodology to
- build spectral libraries of animal feeds. Journal of Near Infrared Spectroscopy 16, 243-248.
- Okparanma, R.N., Mouazen, A.M. 2013. Visible and Near-Infrared Spectroscopy Analysis of a
- Polycyclic Aromatic Hidrocarbon in Soils. The Scientific World Journal.
- Volkers K.C., Wachendorf M., Loges R., Jovanovic N.J., Taube F. 2003. Prediction of the quality
- of forage maize by near-infrared reflectance spectroscopy. Anim. Feed Sci. Technol. Vol.
- 343 109, Issues 1-4, Pages 183–194.
- 344 Hermida, M., Lois, A., Rodriguez-Otero, J.L. 2005. Analysis of nitrogen fractions in silage by
- near-infrared spectroscopy. J. Agric. Food Chem. 53, 1374–1378.
- Nie, Z., Han, J, Liu T., Liu, X. 2008. Application of support vector machine method in prediction
- of lucerne protein fractions by near infrared reflectance spectroscopy. J. Dairy Sci. 91, 2361–
- 348 2369.
- Huang, H., Yu, H., Xu, H., Ying, Y. 2008. Near infrared spectroscopy for on/in-line monitoring
- of quality in foods and beverages: a review, J. Food Eng., 87, 303–13.
- van Barneveld, R.J., Graham, H., Diffey, S. 2018. Predicting the nutritional quality of feed
- ingredients for pigs using near-infrared spectroscopy (NIRS) and chemical analysis. Animal
- 353 Production Science, 58, 709-7018.
- 354 Yan, H., Siesler, H.W. 2018. Hand-held near infrared spectrometers: State of the art
- instrumentation and practical applications. NIR news, 29, 7, 8-12.
- Olakunle, E., Abdul, Rahman T., Orikumhi, I., Yen Leow, C., MHD Nour Hindia. 2018. An
- Overview of Internet of Things (IoT) and Data Analytics in Agriculture: Benefits and
- 358 Challenges. 3758 3773.
- Liu, S., Guo L., Webb, H., Ya, X., Chang, X., 2019. Internet of Things Monitoring System of
- Modern Eco-Agriculture Based on Cloud Computing, 7, 37050-37058.
- 361 Crocombe, R.A. 2018. Portable Spectroscopy. Applied Spectroscopy, 72, 1701–1751.

Rateni, G., Dario, P., Cavallo, F. 2017. Smartphone-Based Food Diagnostic Technologies: A 362 Review. Sensors. 17, 1453, 1-22. 363 Wang, W., Paliwal, 2007. J. Near-infrared spectroscopy and imaging in food quality and safety. 364 Sens. Instrum. food Qual. Saf. 1, 4, 193-207. 365 Texas Instruments, 2016. DLP Technology for Spectroscopy. White Paper DLPA048A. 366 Texas Instruments. 2017. DLP NIRscan Nano EVM User's Guide. 2017. User's Guide. 367 368 Van Soest, P.J., Robertson, J.B., Lewis, B.A., 1991. Methods for Dietary Fiber. Neutral Detergent 369 Fiber, and Nonstarch Polysaccharides in Relation to Animal Nutrition, J. Dairy Sci., 74, 3583 Burns, D.A., Ciurczak, E.W., 2008. Handbook of Near-Infrared Analysis Third Edition, CRC 370 371 Press Taylor & Francis Group 3597. 372 Bertrand, D., Dufour, E. 2000. Infrared Spectroscopy and its analytical applications. Editions 373 TEC & DOC, Paris, France. 374 Barnes, R.J., Dhanoa, M.S., Lister, S.J. 1989. Standard normal variate transformation and de-375 trending of near-infrared diffuse reflectance spectra. Appl. Spectrosc. 43, 772–777. Næs, T., Isaksson, T., Fearn, T., Davies, T. 2004. A User-Friendly Guide to Multivariate 376 377 Calibration and Classification. NIR Publications, Charlton, Chichester, UK. 46, 1, 108–110. Martens, H., Naes, T. 1989. Multivariate calibration. John Wiley and Sons Inc., Chichester, UK. 378 379 Wold, S., Sjöström,, M., Eriksson L. 2001. PLS-regression: a basic tool of chemometrics. 380 Chemometrics. Intell. Lab. Syst. 58, 2, 109–130. 381 382 383 384 385 386

388 FIGURES

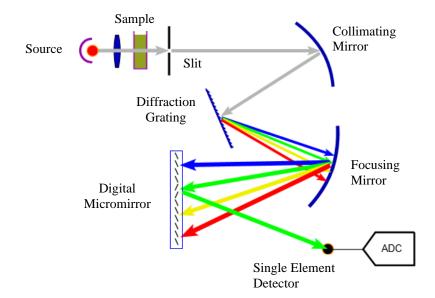
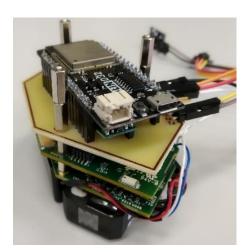


Fig. 1. Spectrometer using a micromirror array and a single element detector.



a)



b)

Fig. 2. (a) NIRscan Nano module and (b) Microcontroller board places over NIRscan module.

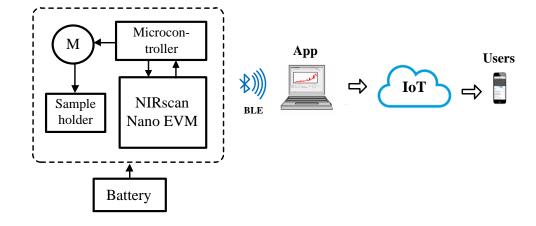
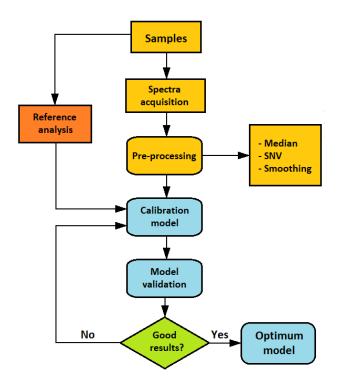


Fig. 3. General scheme of the proposed measurement system (M = servomotor).





Fig. 4. NIRS system showing the sample holder (a) without alfalfa and (b) sample holder with alfalfa.



419 Fig. 5. Process to obtain the chemometric model.

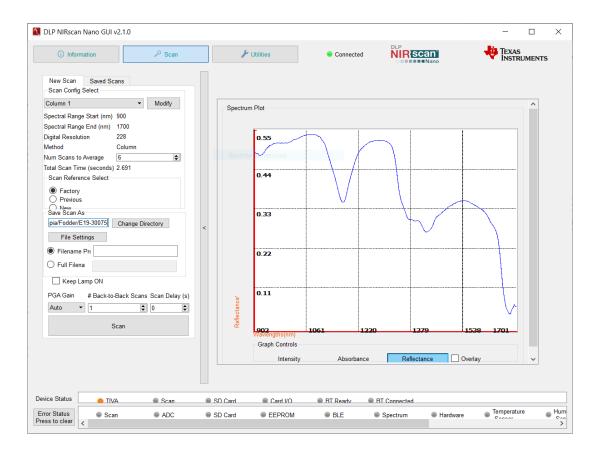


Fig. 6. NIRscan Nano GUI scan screen.

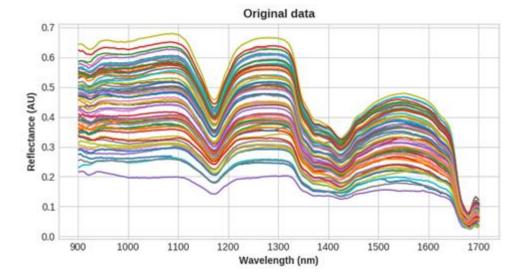


Fig. 7. Spectra of alfalfa samples (N = 90).

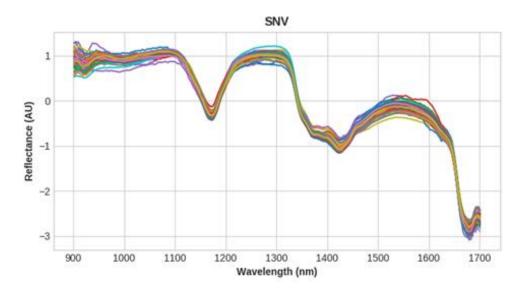


Fig.8. Spectra after applying SNV correction to alfalfa samples.

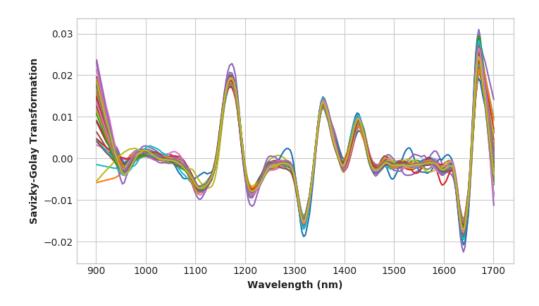


Fig. 9. Savitzky–Golay filter applied to alfalfa sample data.

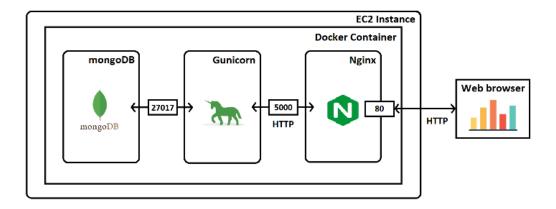


Fig. 10. Dashboard structure on the cloud.

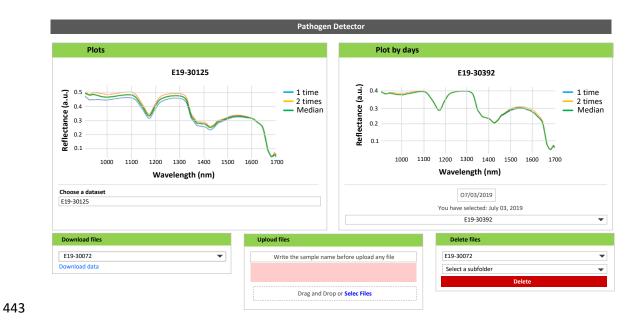


Fig. 11. Dashboard for user visualization.